

# Modelling secondary AML using human induced pluripotent stem cells

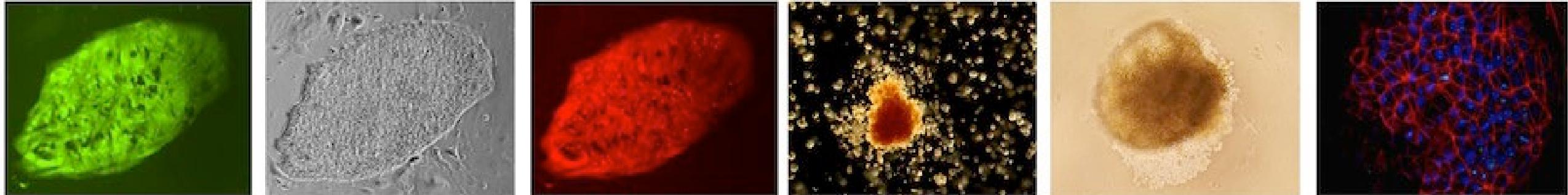


Eirini Papapetrou, MD, PhD

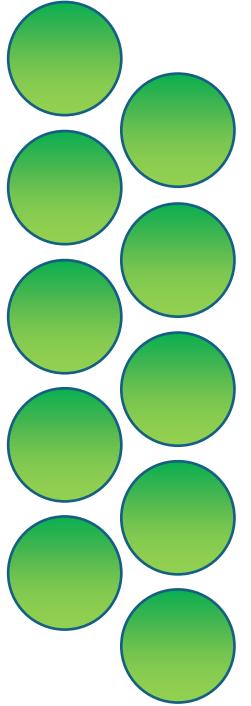
Icahn School of Medicine at Mount Sinai



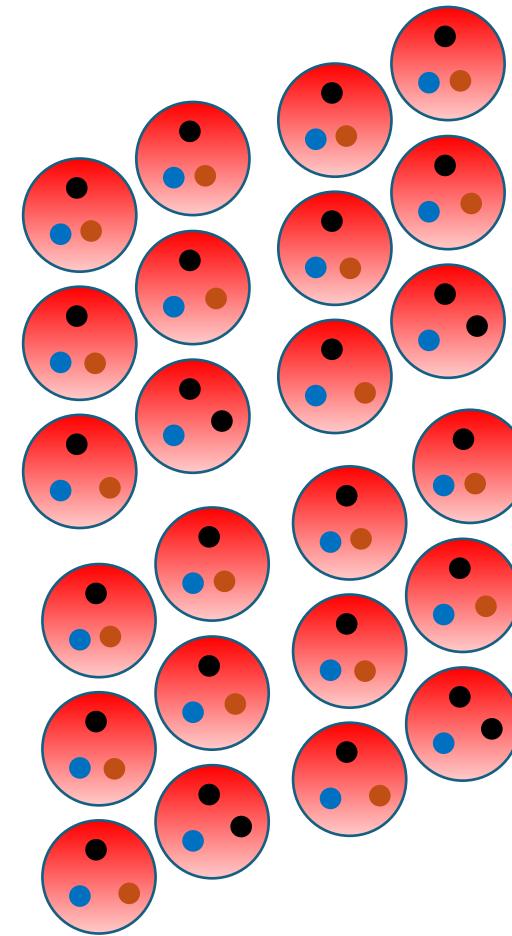
Center for Advancement  
of **Blood Cancer Therapies**



# How does a normal HSPC turn bad?

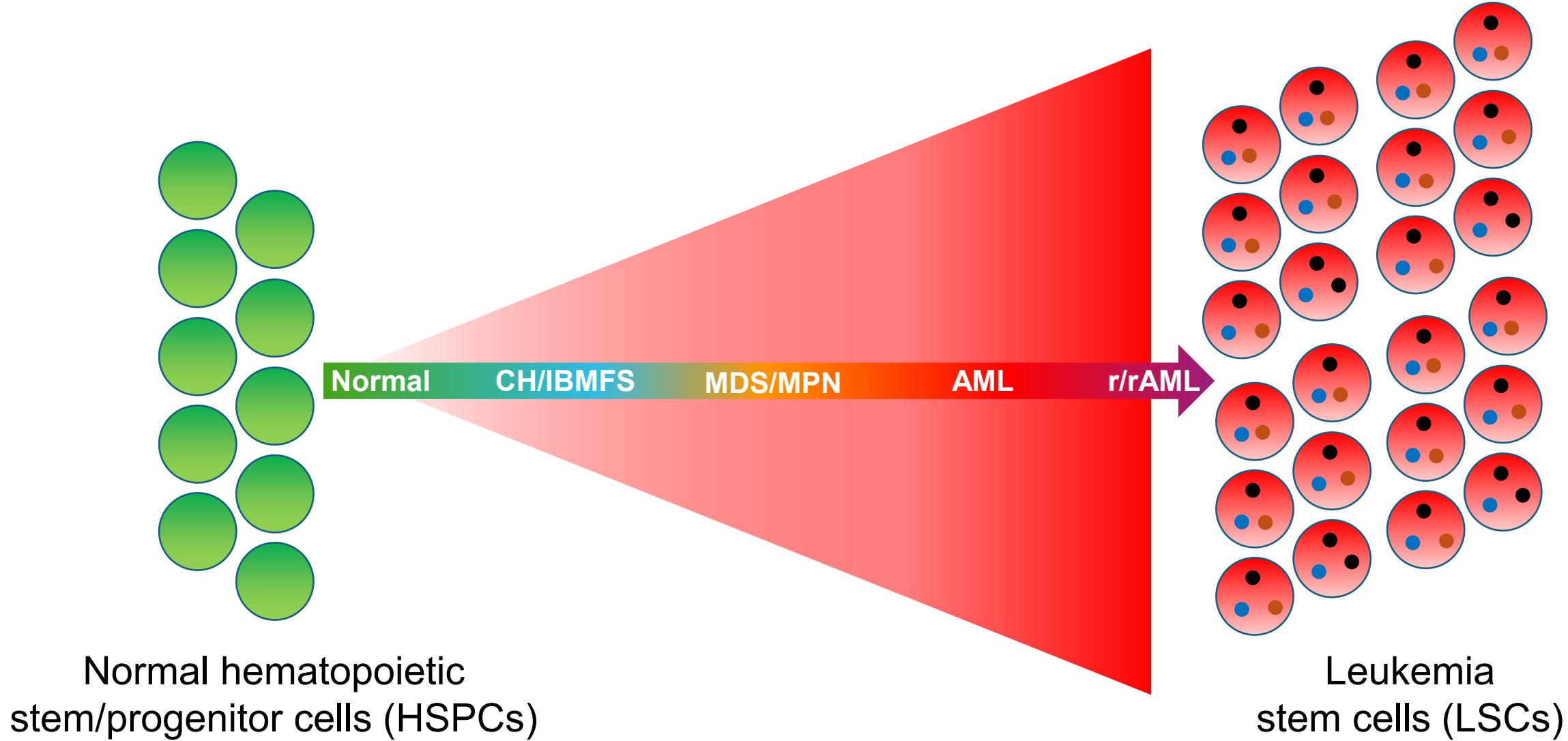


Normal hematopoietic  
stem/progenitor cells (HSPCs)



Leukemia  
stem cells (LSCs)

# How does a normal HSPC turn bad?

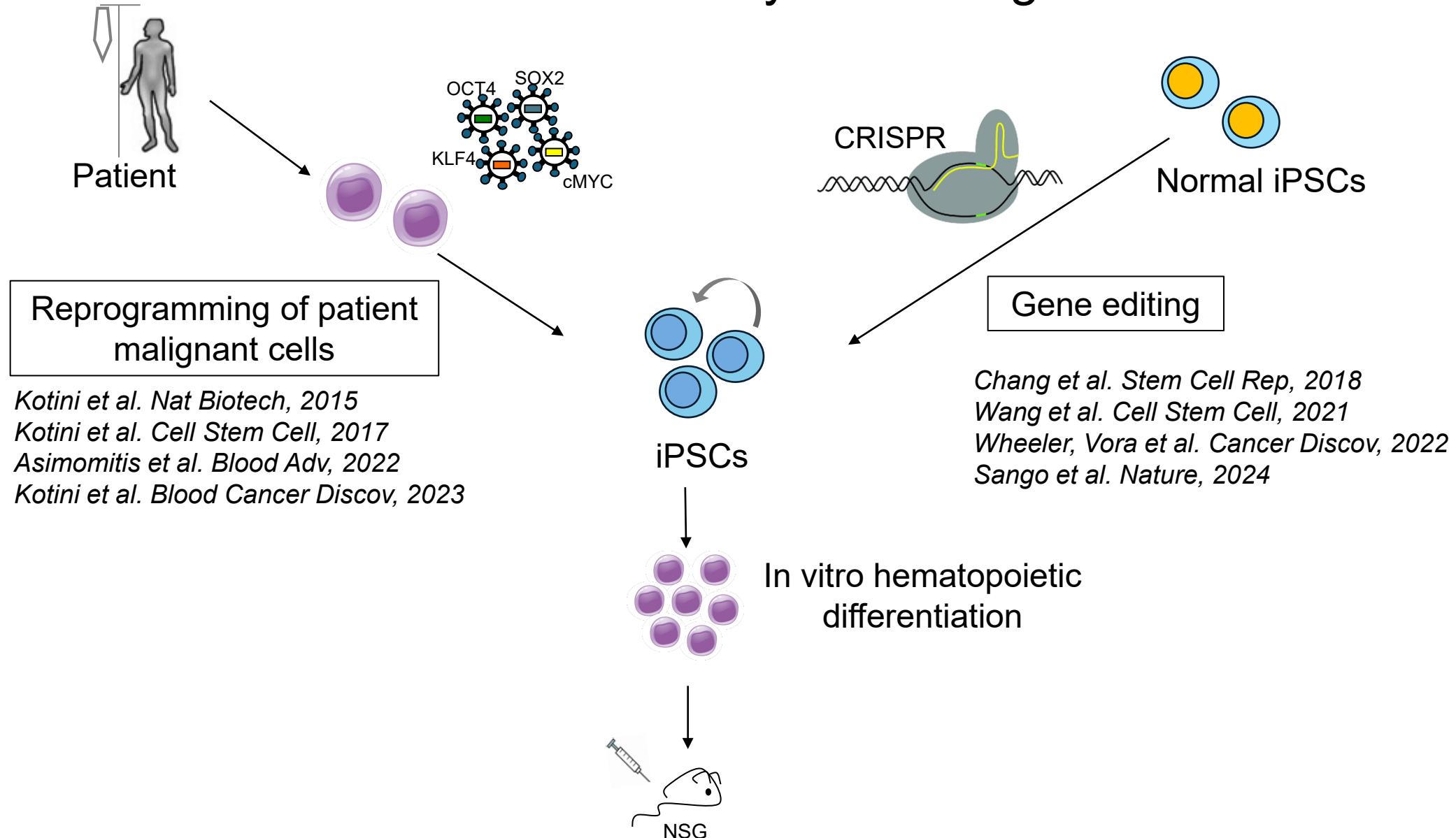




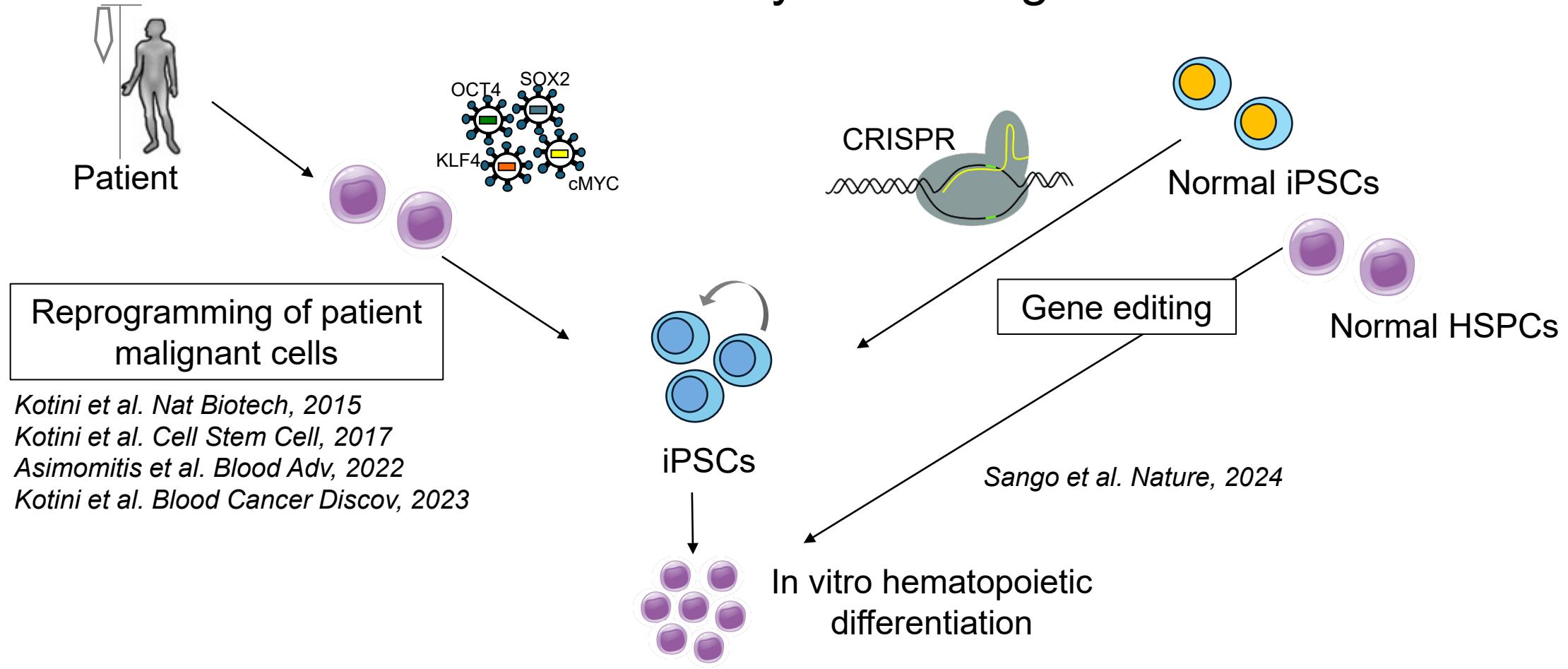
Richard Feynman  
1918-1988

*“What I cannot create I do not understand”*

# Human iPSC models of myeloid malignancies



# Human iPSC models of myeloid malignancies



## Key advantages:

- Human
- Genetically faithful models
- Isogenic
- Unlimited cell numbers, high throughput
- Functional studies

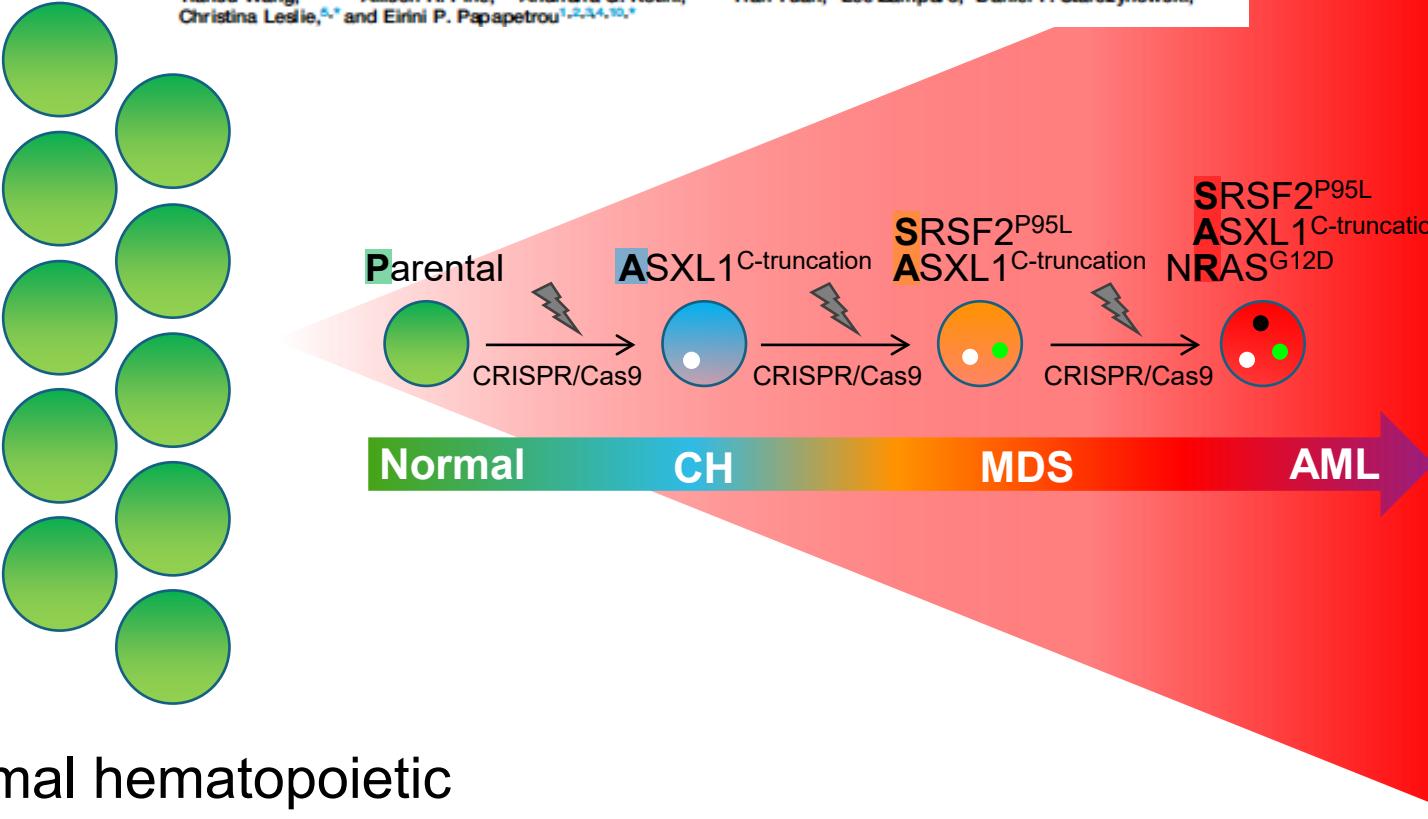
# A synthetic model of CH-MDS-sAML progression

Cell Stem Cell

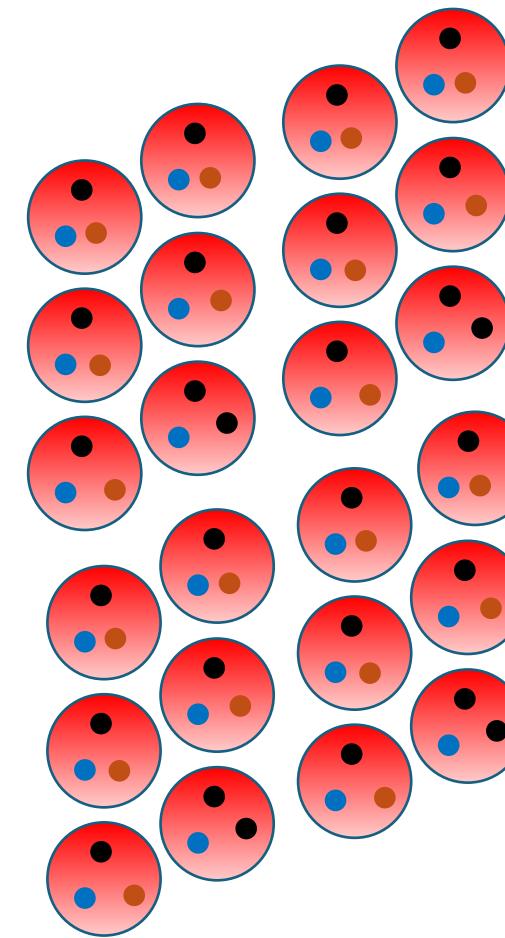
Article

Sequential CRISPR gene editing in human iPSCs charts the clonal evolution of myeloid leukemia and identifies early disease targets

Tiansu Wang,<sup>1,2,7,4,9</sup> Allison R. Pine,<sup>5,9</sup> Andriana G. Kotini,<sup>1,2,3,4</sup> Han Yuan,<sup>5</sup> Lee Zamparo,<sup>5</sup> Daniel T. Starczynowski,<sup>6,7,8</sup> Christina Leslie,<sup>5,\*</sup> and Eirini P. Papapetrou<sup>1,2,3,4,10,\*</sup>



Normal hematopoietic  
stem/progenitor cells (HSPCs)



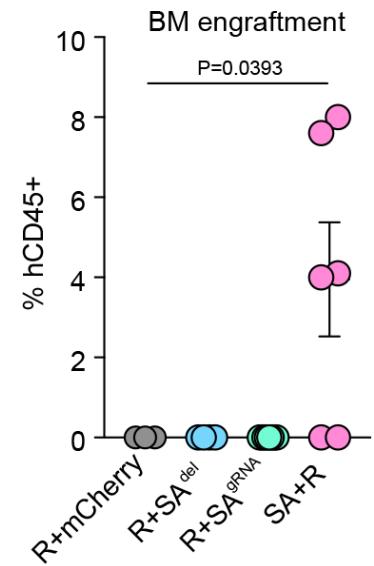
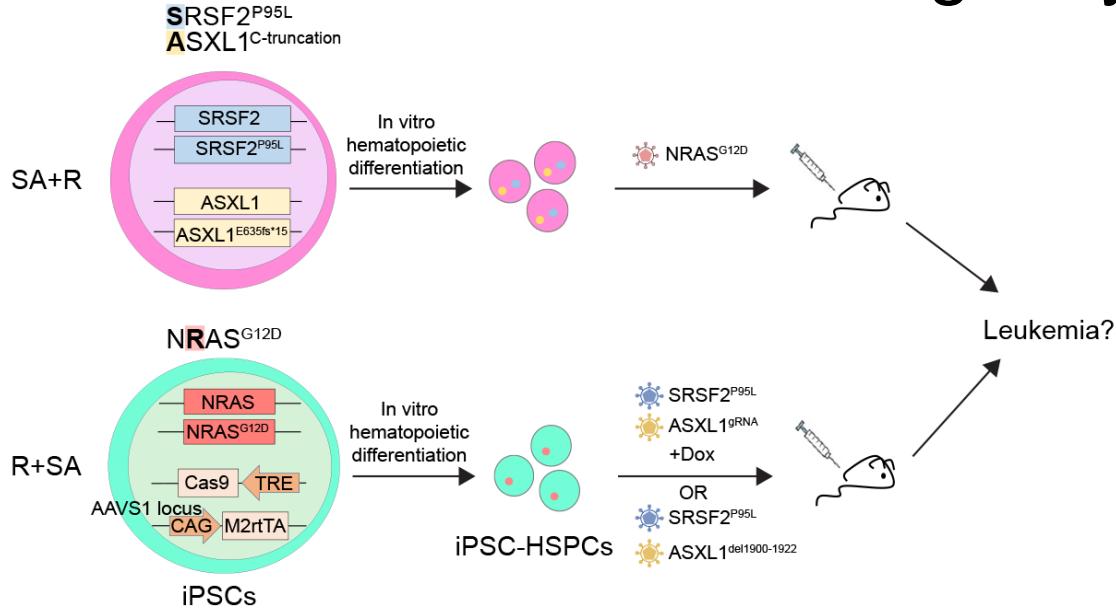
Leukemia  
stem cells (LSCs)

# Why are RAS mutations always late in AML?

*N*/KRAS mutations:

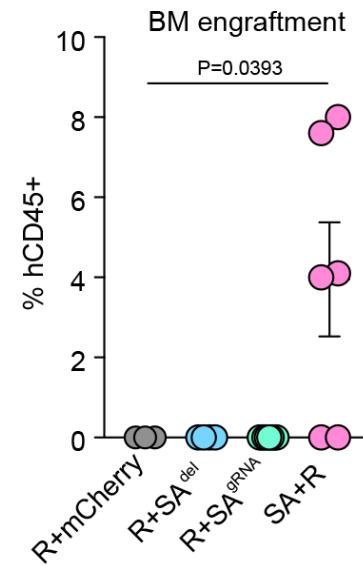
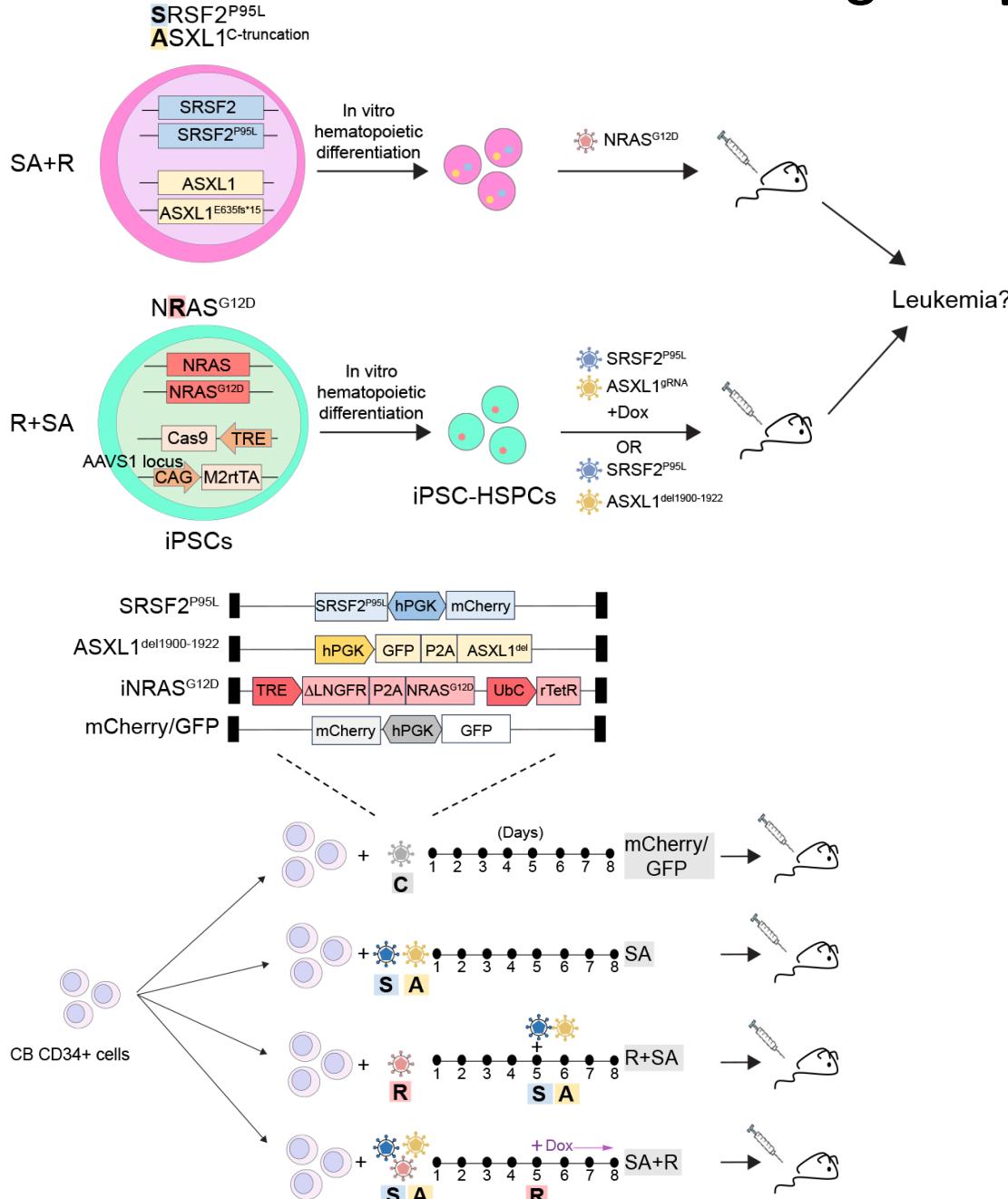
- are present in up to 30% of AML patients
- are almost always subclonal
- are often acquired upon relapse
- are often acquired during progression from MDS to sAML
- are, in contrast, early truncal events in epithelial cancers

# $\text{NRAS}^{\text{G12D}}$ is an obligatory late mutation

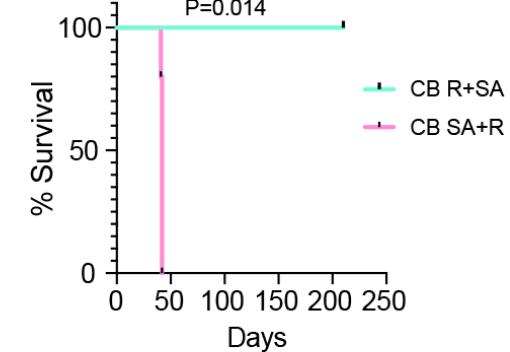
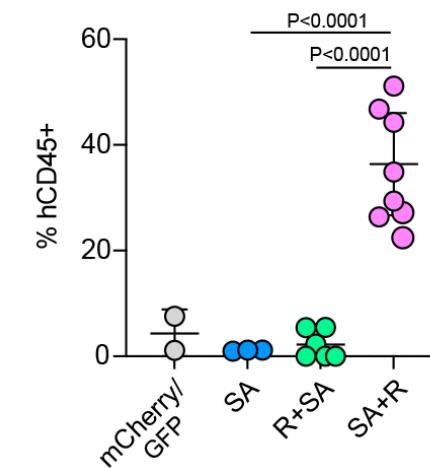


Junya Sango

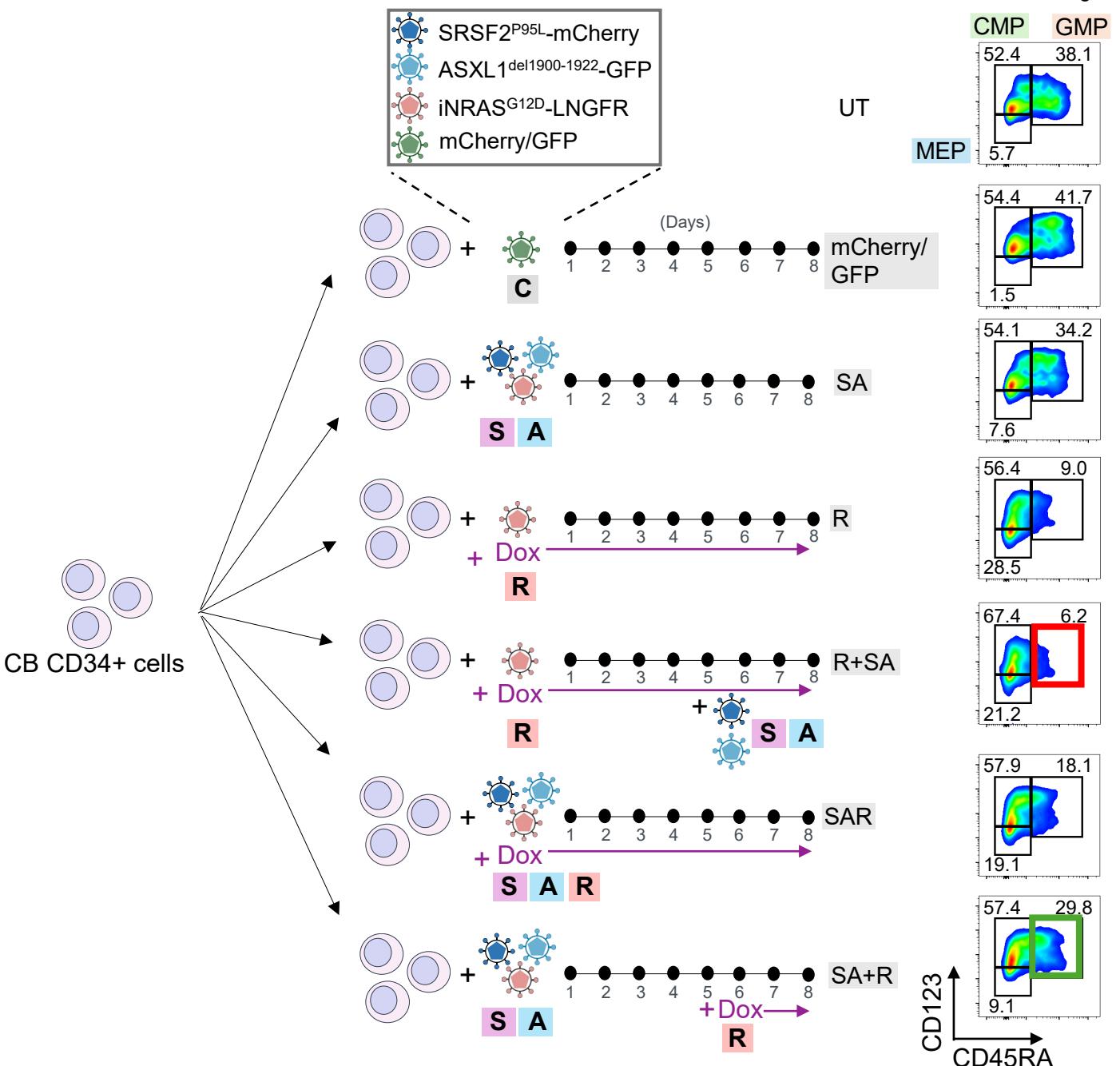
# NRAS<sup>G12D</sup> is an obligatory late mutation



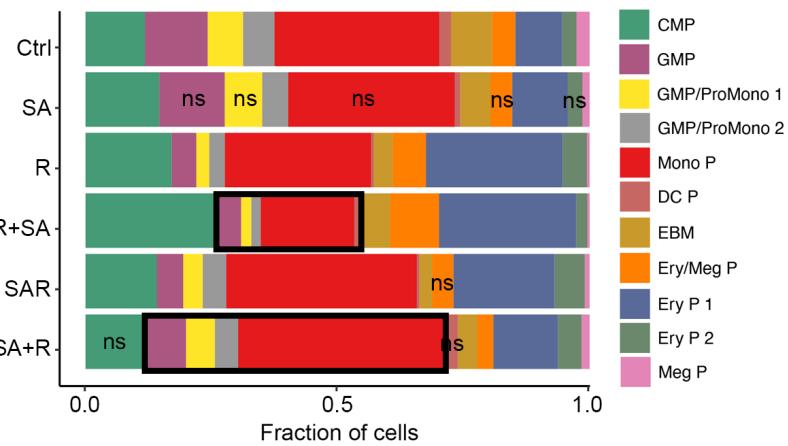
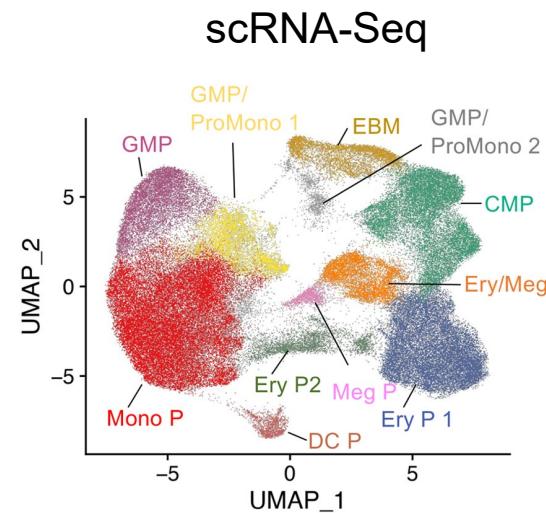
Junya Sango



# RAS mutations transform GMPs harboring preexisting mutations

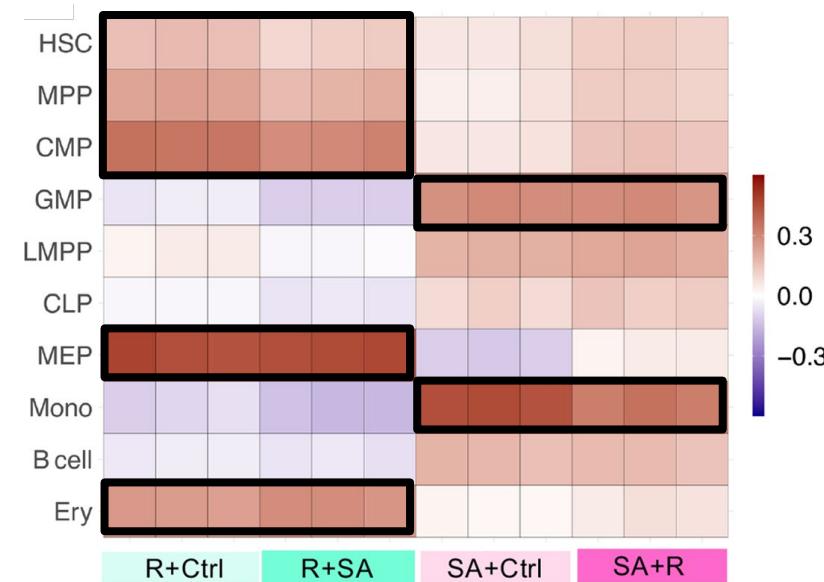


# RAS mutations transform GMPs harboring preexisting mutations

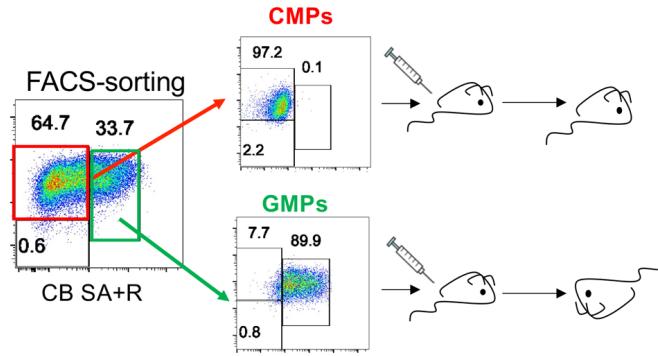


Saul Carcamo Lewis Tomalin Gulay Ulukaya

## ATAC-Seq

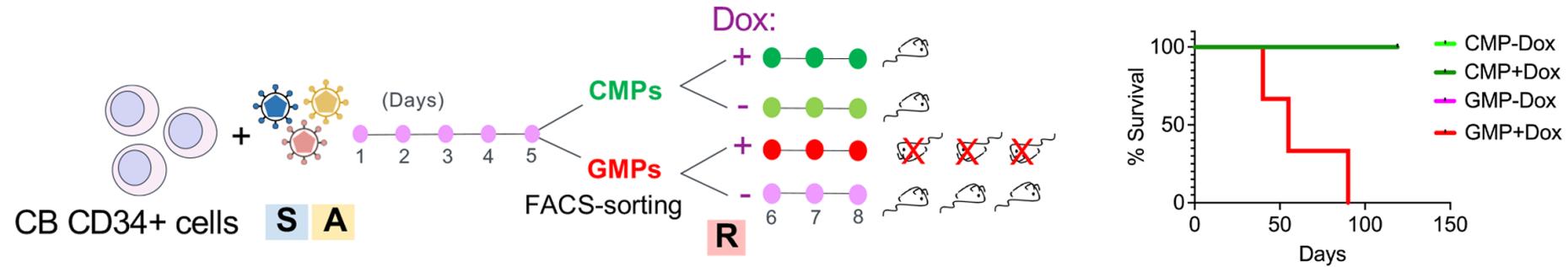


# RAS mutations transform GMPs harboring preexisting mutations



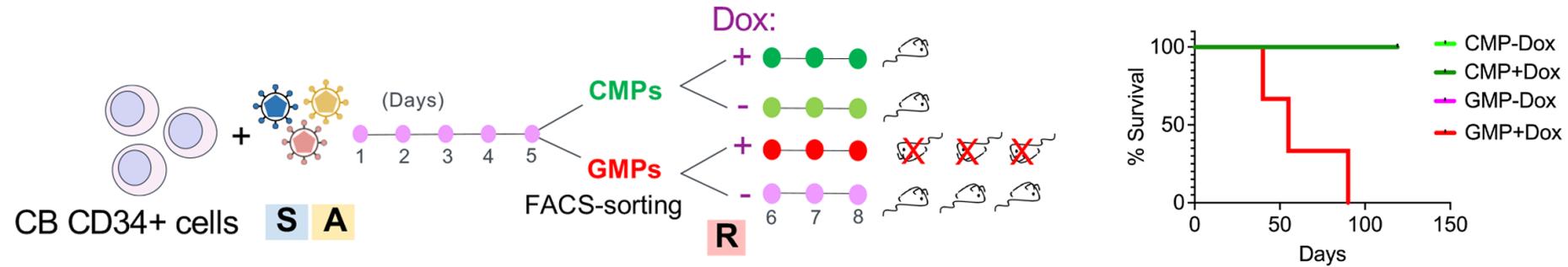
➤ SA+R GMPs are the leukemia initiating cells

# RAS mutations transform GMPs harboring preexisting mutations

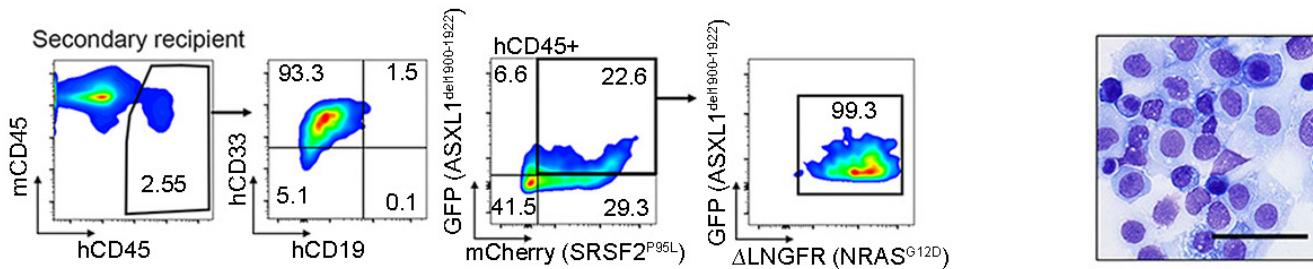


➤ SA GMPs are the target cells of transformation by RAS mutations

# RAS mutations transform GMPs harboring preexisting mutations

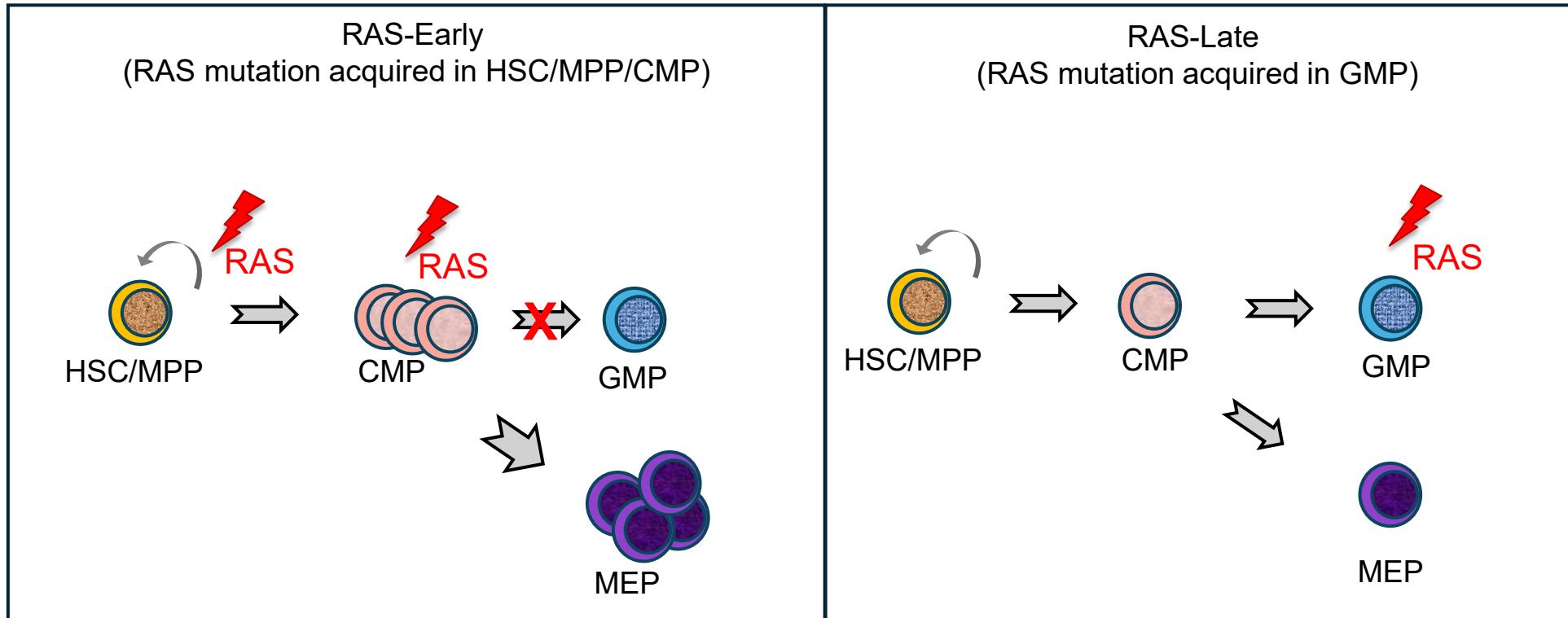


- SA GMPs are the target cells of transformation by RAS mutations

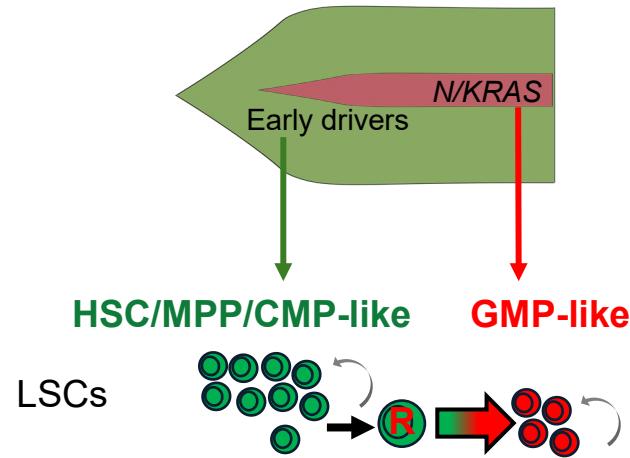


- SA+R GMPs are leukemia stem cells

# RAS mutations transform GMPs harboring preexisting mutations



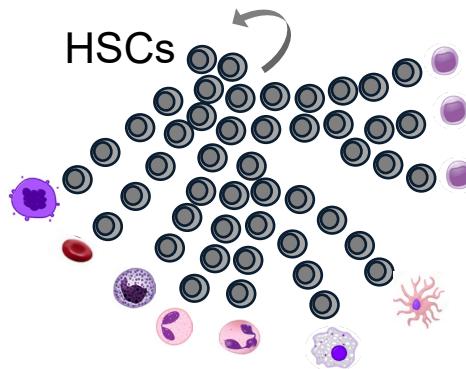
- RAS mutations transform progenitors committed to the myelomonocytic lineage (GMPs) that have previously acquired driver mutations, i.e. are descendants of an ancestral clone originating from a more primitive (HSC/MPP/CMP) cell



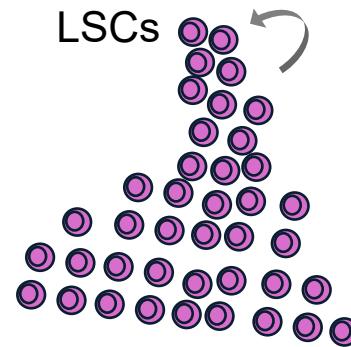
# AML hierarchies and phenotype

FAB CLASSIFICATION	
	<b>M0:</b> Undifferentiated acute myeloblastic leukemia (5%)
	<b>M1:</b> Greater number of myeloblasts with <10% granulocytic differentiation.
	<b>M2:</b> Myeloblasts in great number with granulocytic differentiation >10%, NSE <20%.
	<b>M3:</b> Promyelocytes that are hyper granular with many Auer rods on CAE or Wright-stain and variant form cells with reniform nuclei, multilobed or bilobed, primeval cells with multiple Auer rods or relative scarcity of Hypergranular promyelocytes.
	<b>M4:</b> >20% but <80% NSE-butrate positivity in Monocytic cells
	<b>M5:</b> Monocytic cells with >80% NSE positivity. (a) Monocytic differentiated (b) Monocytic, differentiated.
	<b>M6:</b> >30% myeloblasts with more than 50% erythroblasts eliminating the erythroid cells.
	<b>M7:</b> Acute megakaryoblastic leukemia <5%

Normal hematopoiesis



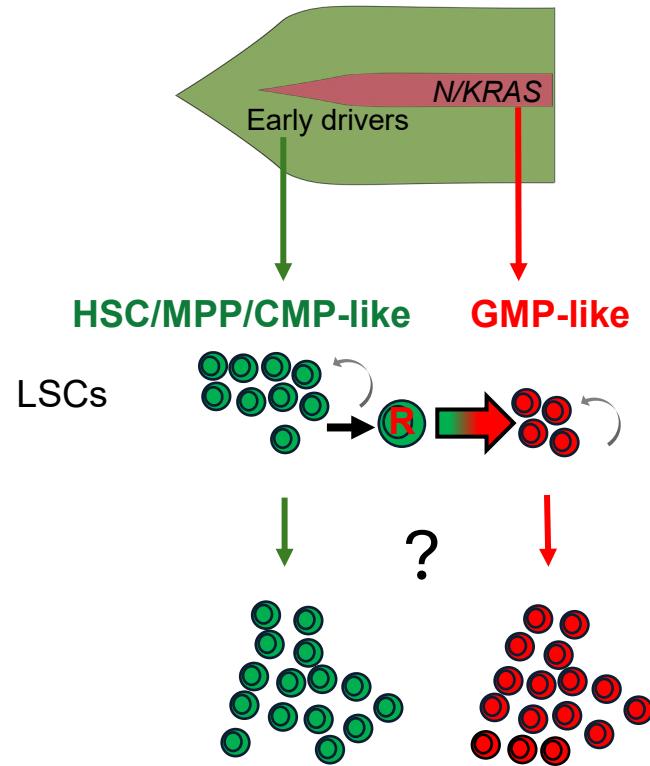
AML with deep hierarchy



AML with shallow hierarchy

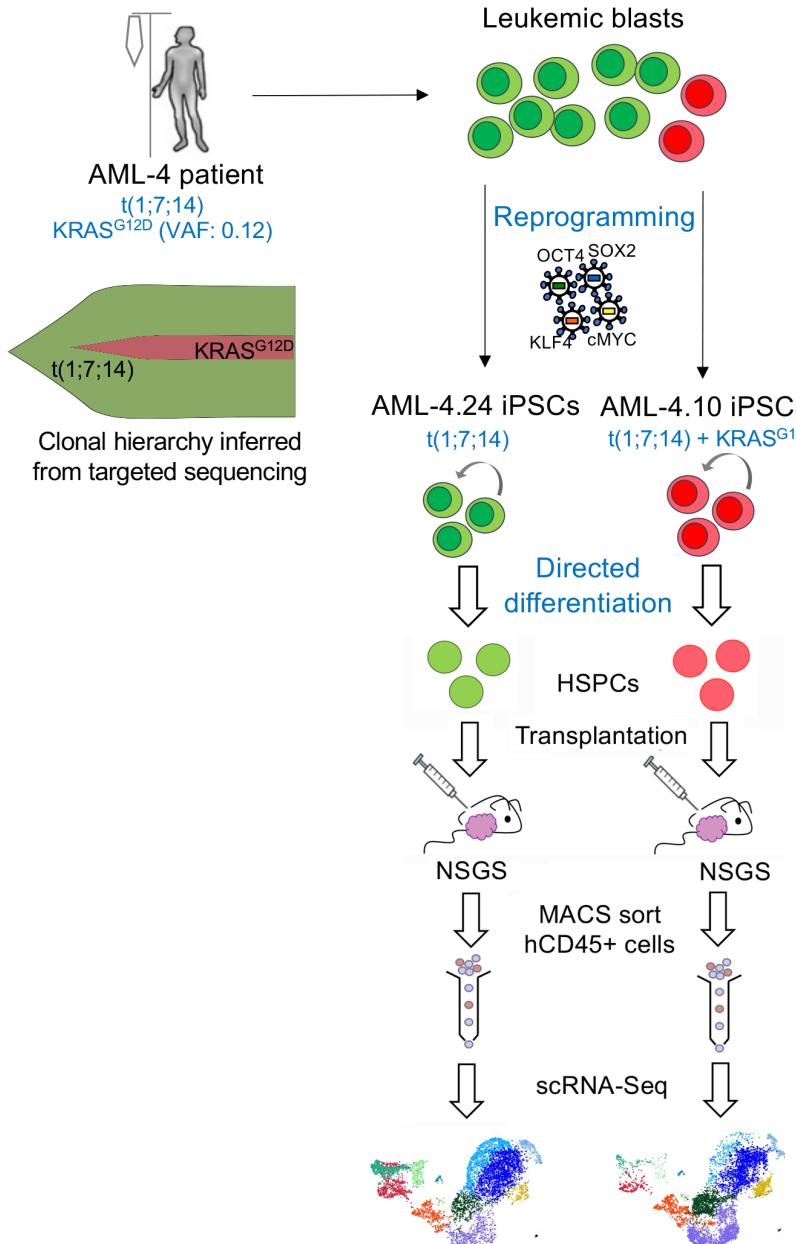


- RAS mutations transform progenitors committed to the myelomonocytic lineage (GMPs) that have previously acquired driver mutations, i.e. are descendants of an ancestral clone originating from a more primitive (HSC/MPP/CMP) cell



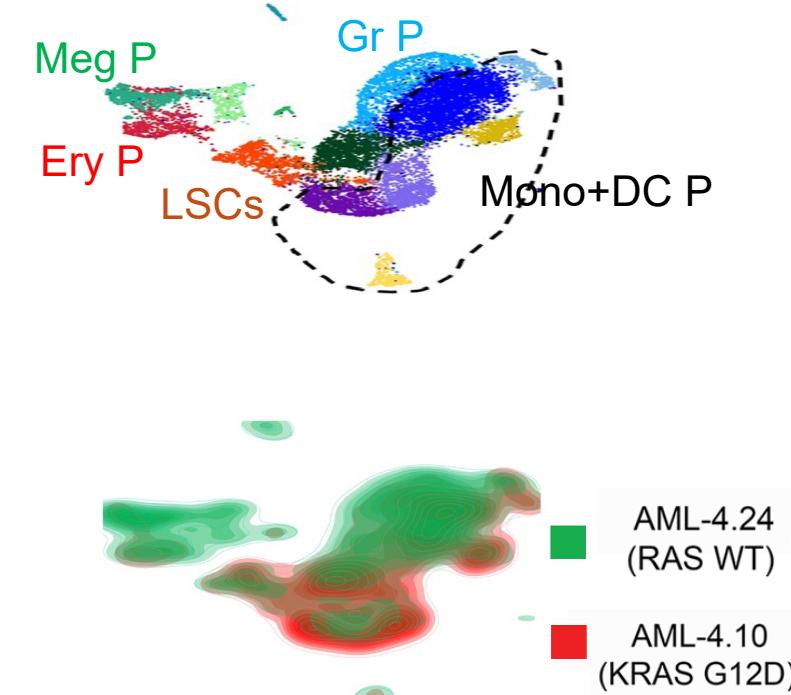
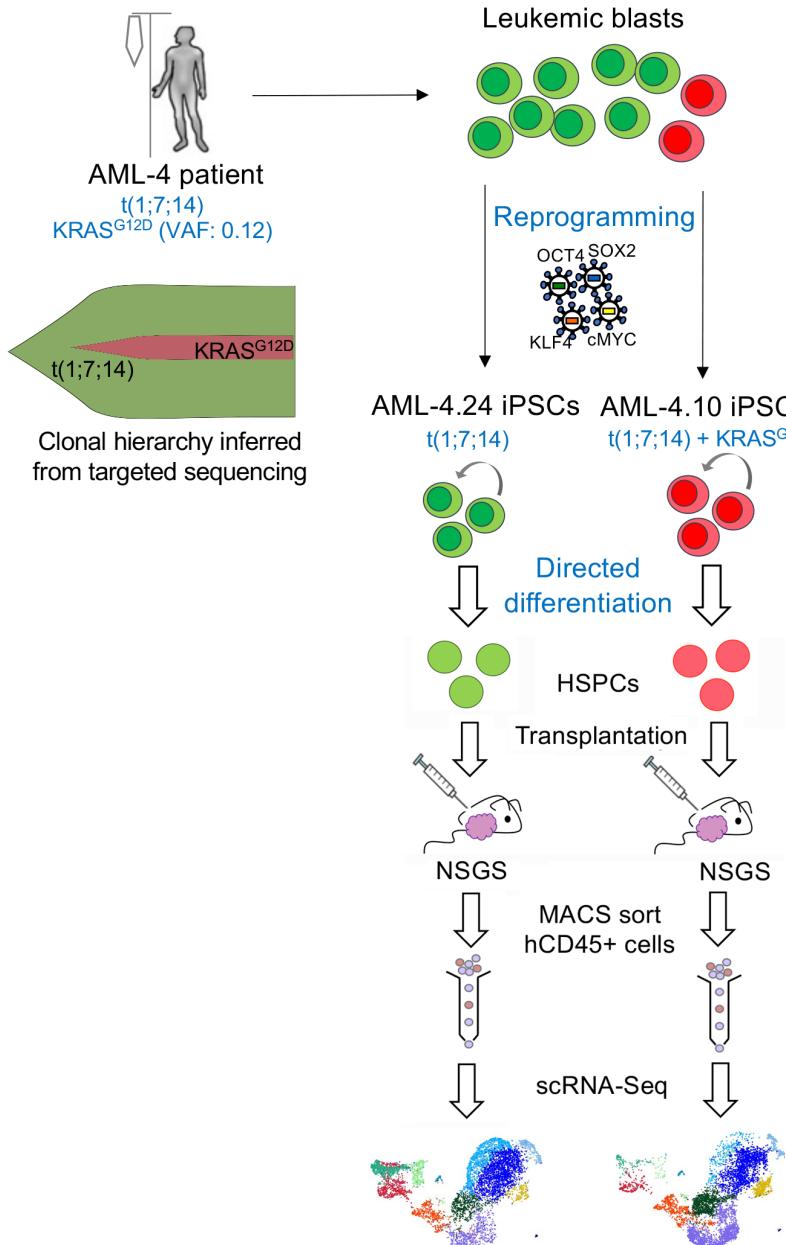
# Comparison of RAS-MT and RAS-WT cells within the same AML patient

## Patient-derived iPSCs



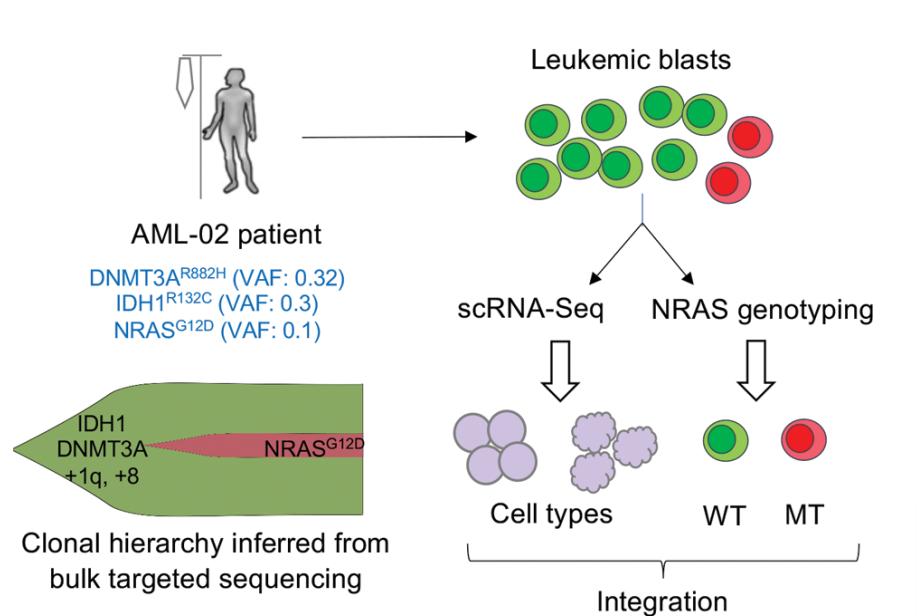
# Comparison of RAS-MT and RAS-WT cells within the same AML patient

## Patient-derived iPSCs



# Comparison of RAS-MT and RAS-WT cells within the same AML patient

## Genotyping of Transcriptomes (GoT)



Maria Sirenko



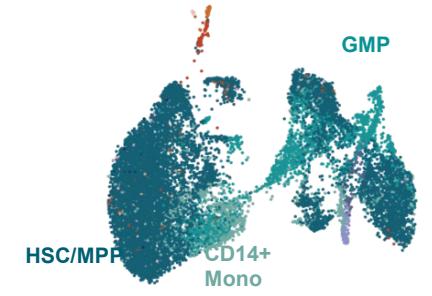
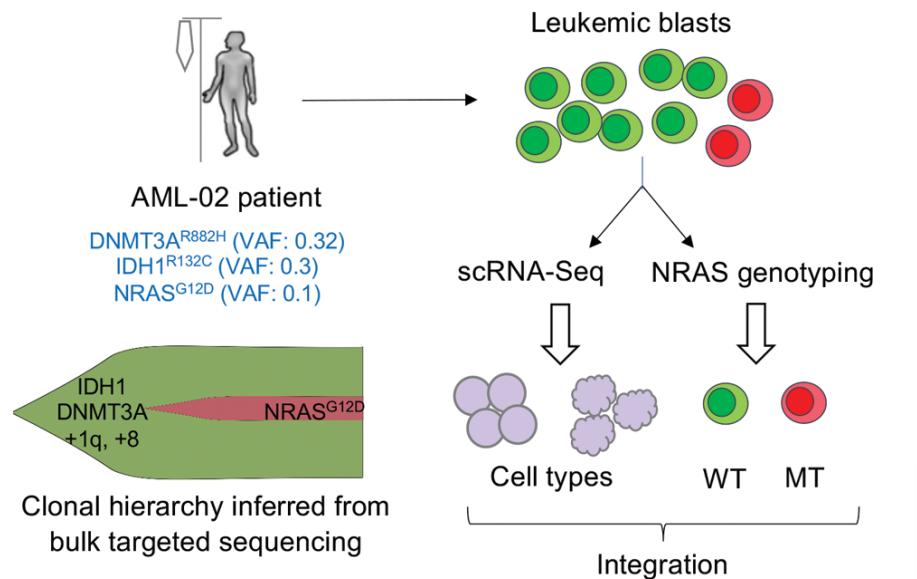
Elli Papaemmanuil



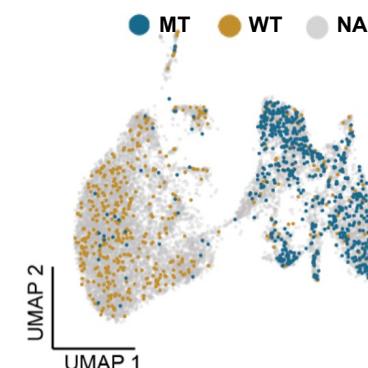
Dan Landau

# Comparison of RAS-MT and RAS-WT cells within the same AML patient

## Genotyping of Transcriptomes (GoT)



NRAS (n= 1184 cells genotyped)



Maria Sirenko

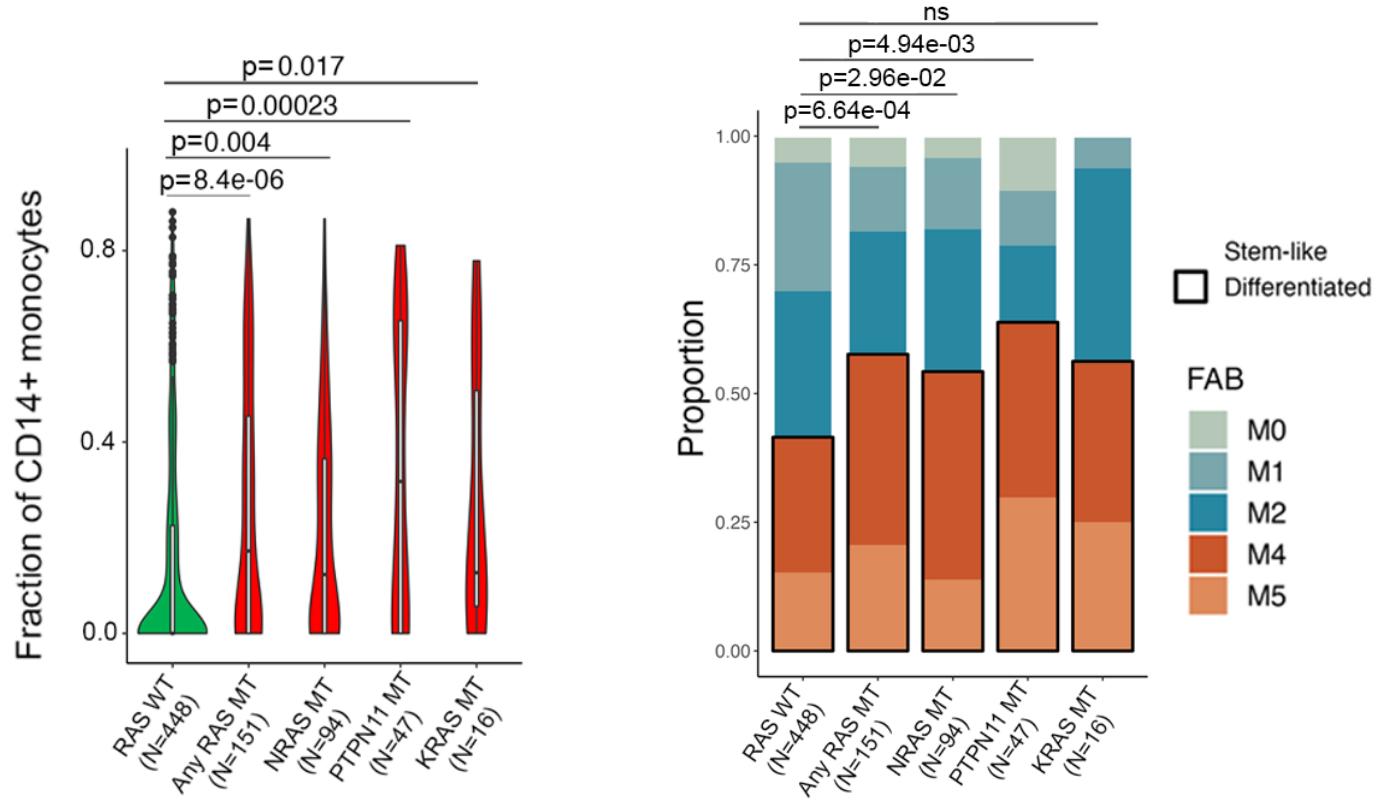


Elli Papaemmanuil



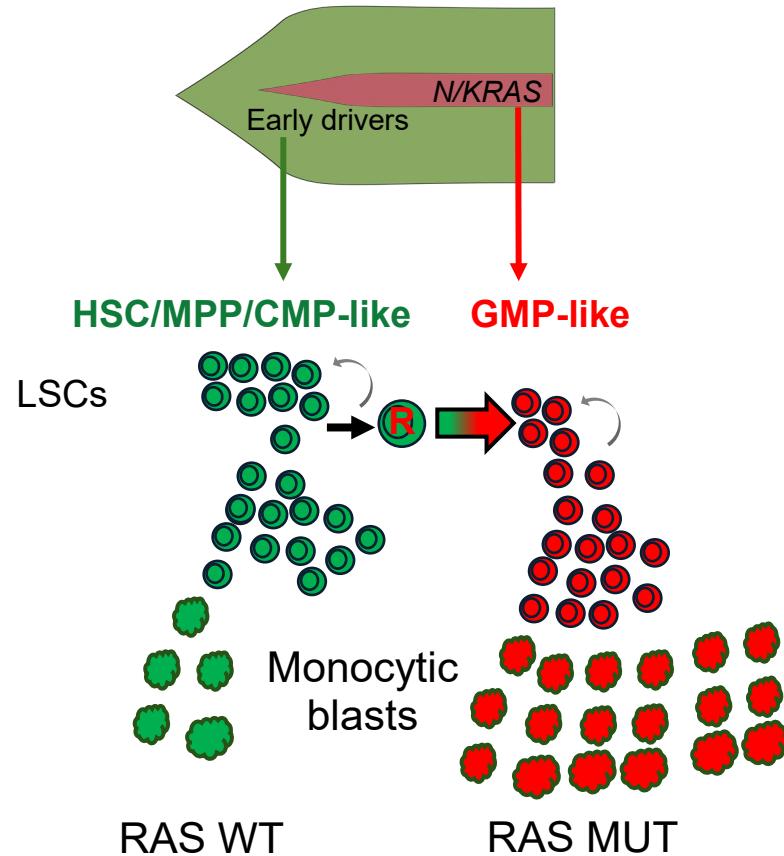
Dan Landau

# RAS-MT AML is monocytic



Maria Sirenko  
Ann-Kathrin Eisfeld  
Bettina Nadorp

- RAS-mutant LSCs give rise to AML blasts with monocytic differentiation, whereas ancestral LSCs generate primitive AML blasts



# Venetoclax (VEN) resistance in AML

? Determinants of relapse/resistance

- VEN resistance/relapse associated with monocytic AML.
  - VEN resistance/relapse associated with RAS pathway mutations.
- RAS pathway mutations cause monocytic AML.

# Venetoclax (VEN) resistance in AML

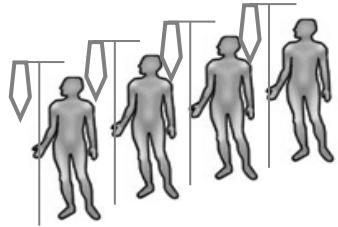
? Determinants of relapse/resistance

- VEN resistance/relapse associated with monocytic AML.
- VEN resistance/relapse associated with RAS pathway mutations.

➤ RAS pathway mutations cause monocytic AML.

➤ Is it the monocytic differentiation stage or the RAS mutational status  
that is the cause of VEN resistance?

# N/KRAS mutation, but not monocytic stage, is associated with poor outcomes in AML patients in a prospective clinical trial of VEN+DEC



VEN  
+  
DEC

Older/unfit patients with  
newly diagnosed AML

NCT03404193



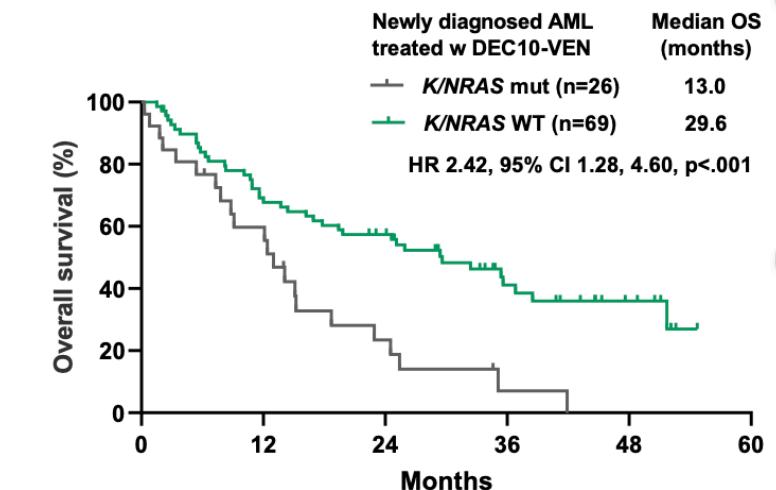
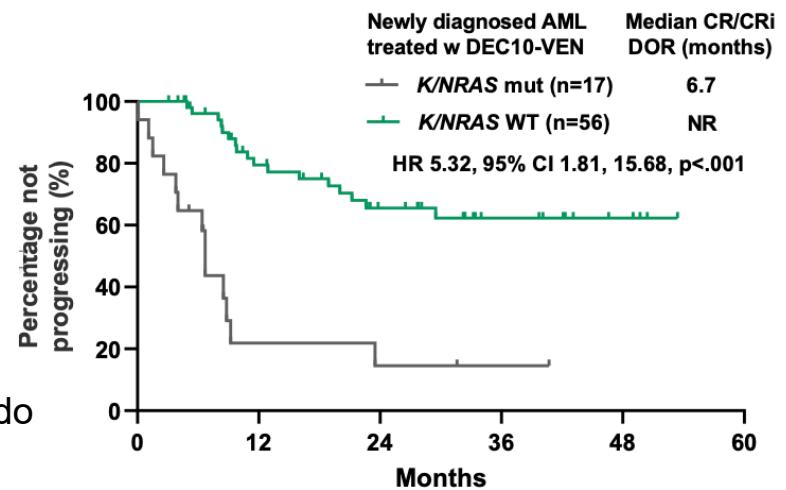
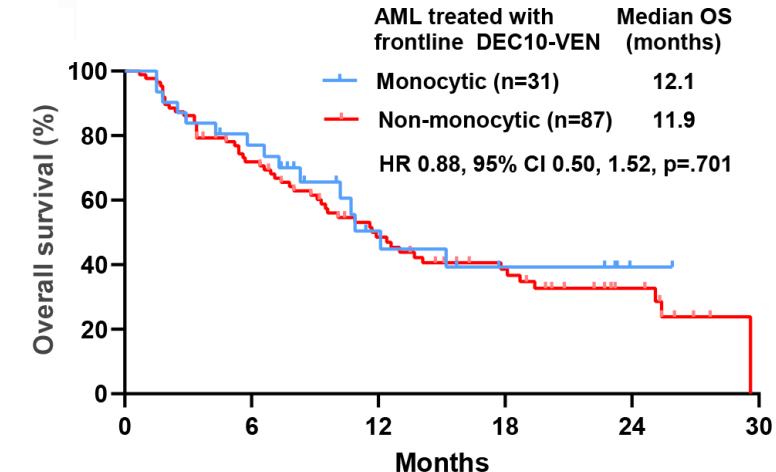
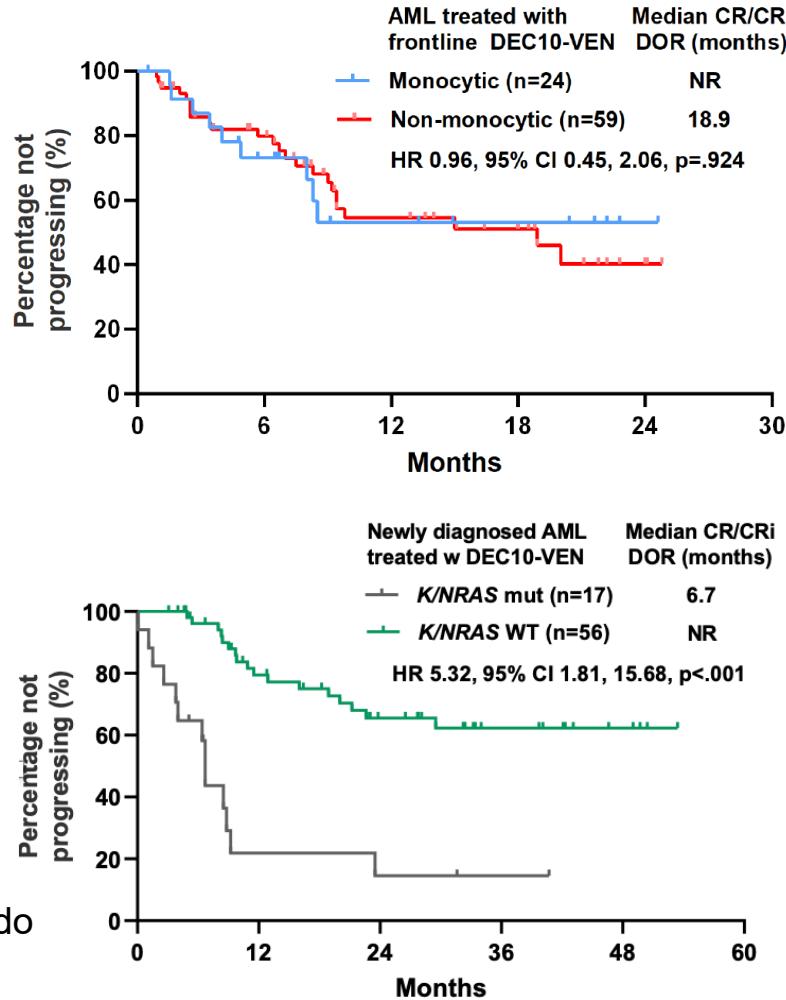
Marina Konopleva



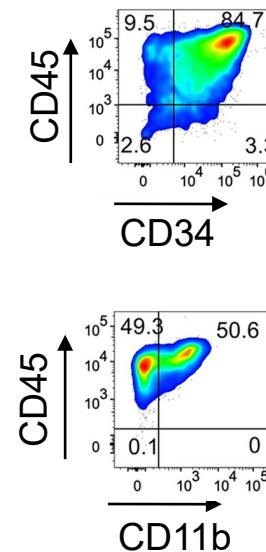
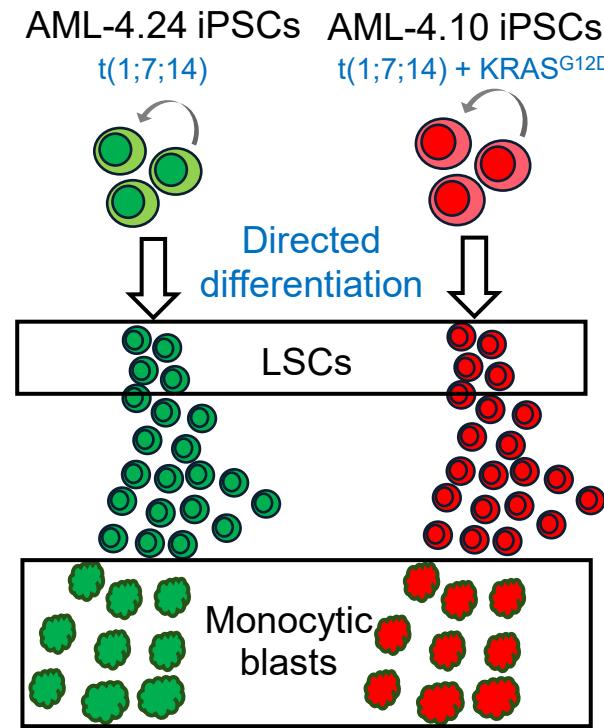
Abhi Maiti



Courtney DiNardo



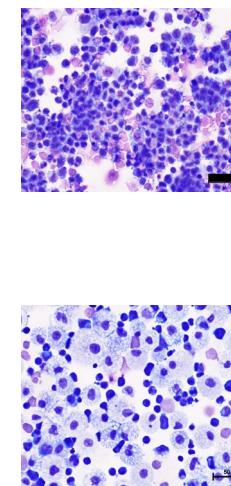
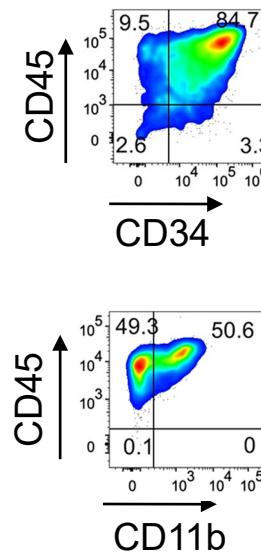
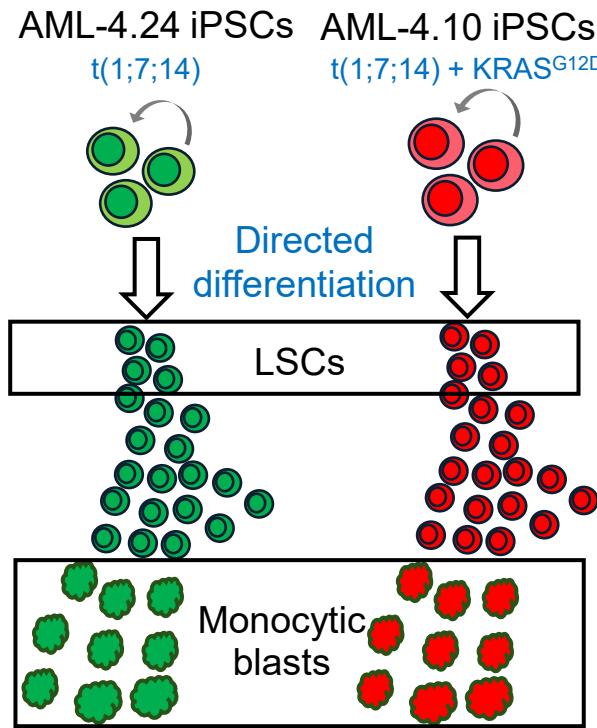
# Separating the impact of differentiation state vs mutational status



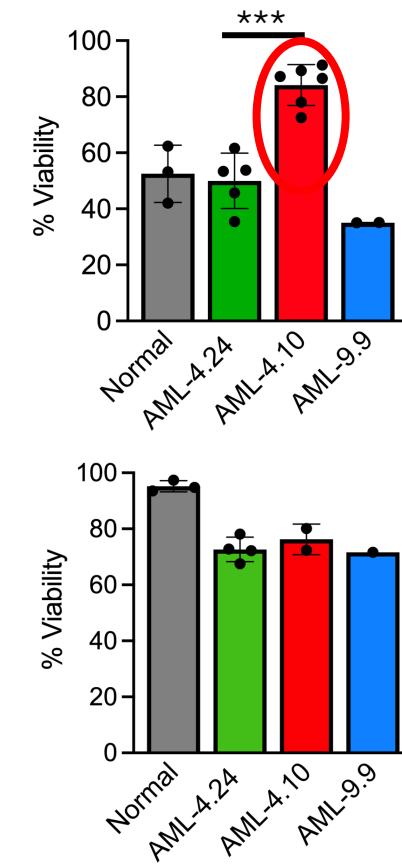
+Ven

+Ven

# RAS-WT LSCs are sensitive to VEN, but RAS-MT LSCs are resistant

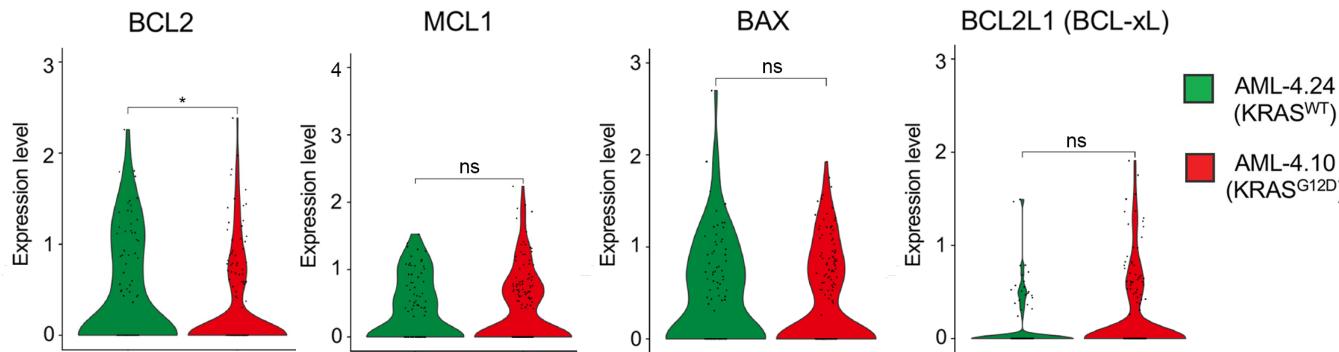


+Ven

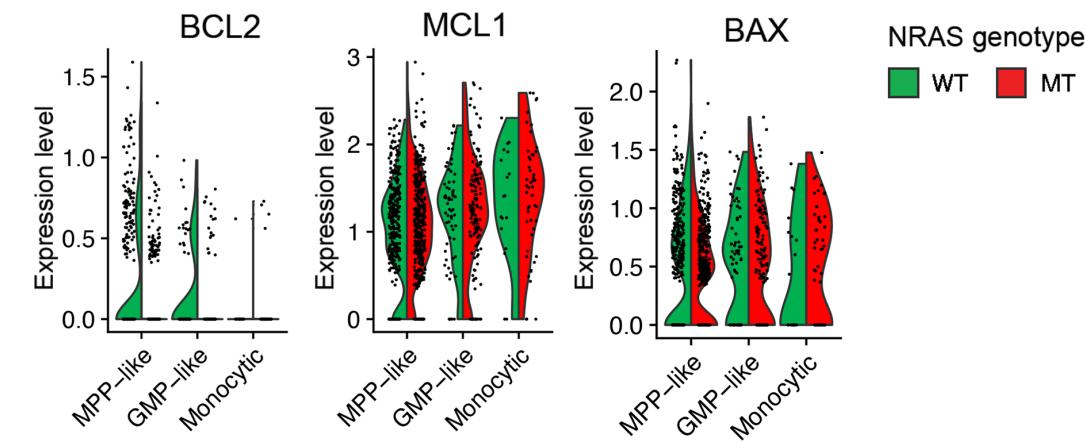


# RAS mutations and not the GMP state confer VEN resistance to LSCs

Patient-derived AML-iPSC xenografts

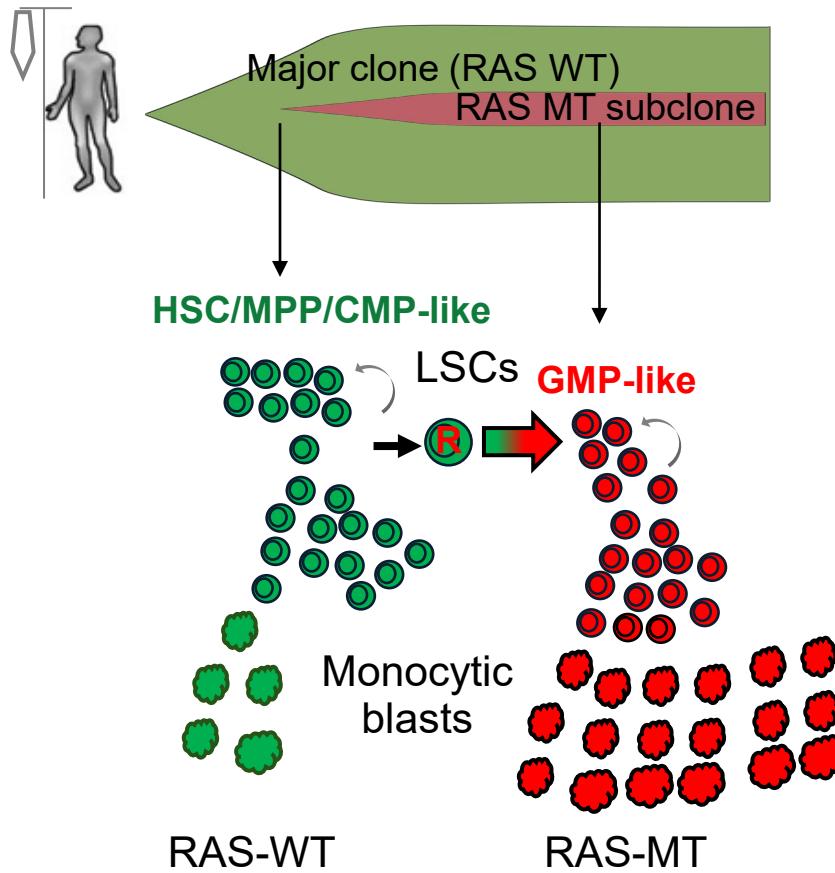


Genotyping of Transcriptomes (GoT)

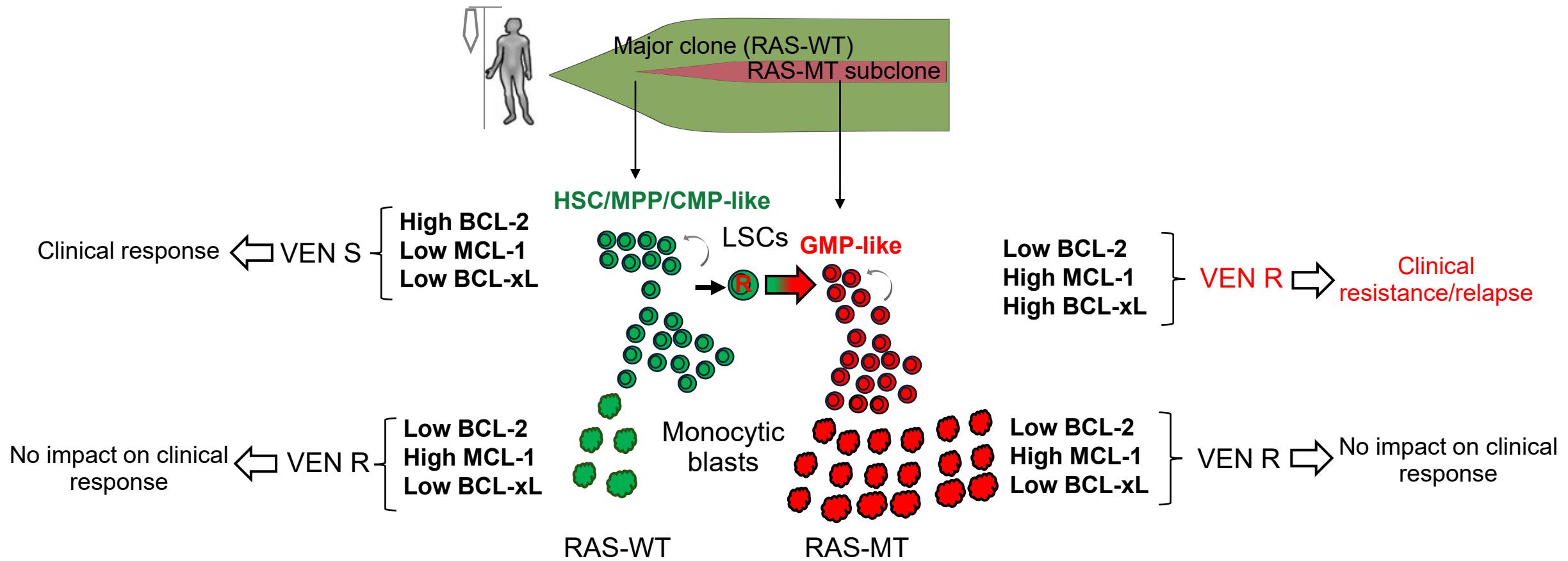


RAS-MT LSCs have ↓ BCL2 and ↑ MCL1 and ↑ BCL-xL

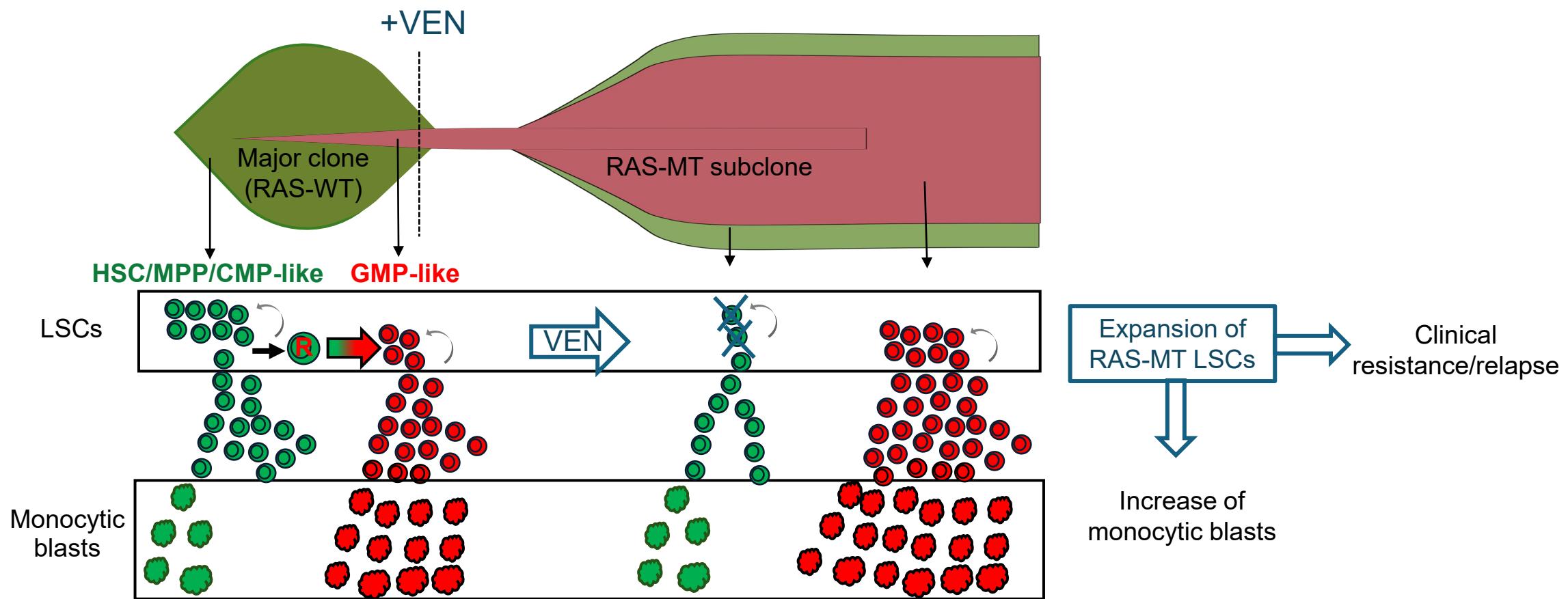
# Unified model of VEN resistance



# Unified model of VEN resistance



# Unified model of VEN resistance



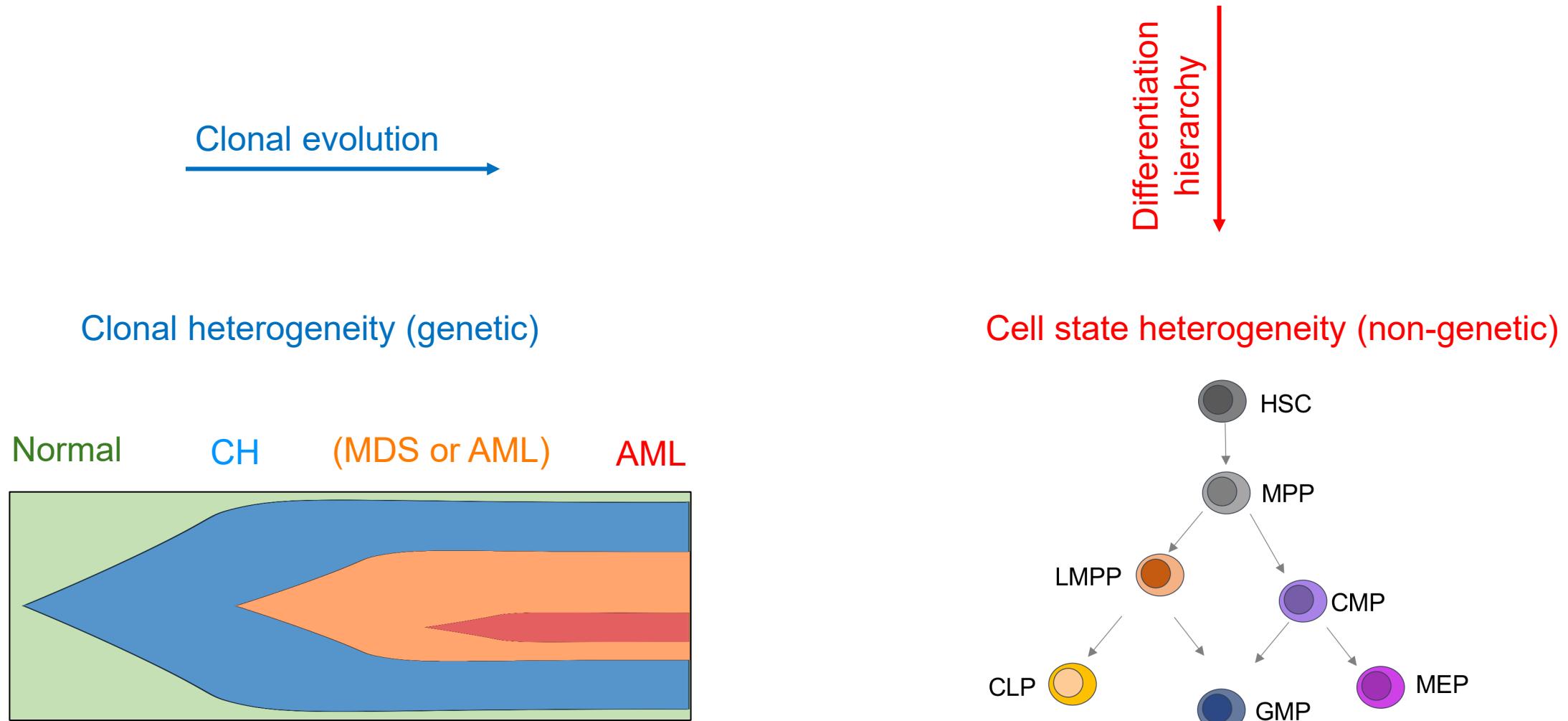
**Article**

# RAS-mutant leukaemia stem cells drive clinical resistance to venetoclax

Junya Sango<sup>1,2,3,4,5,22</sup>, Saul Carcamo<sup>1,2,3,4,5,6,22</sup>, Maria Sirenko<sup>7,8,22</sup>, Abhishek Maiti<sup>9,22</sup>, Hager Mansour<sup>1,2,3,4,5</sup>, Gulay Ulukaya<sup>1,2,3,4,5,6</sup>, Lewis E. Tomalin<sup>2,3,4,5,6</sup>, Nataly Cruz-Rodriguez<sup>1,2,3,4,5</sup>, Tiansu Wang<sup>1,2,3,4,5</sup>, Małgorzata Olszewska<sup>1,2,3,4,5</sup>, Emmanuel Olivier<sup>1,2,3,4,5</sup>, Manon Jaud<sup>1,2,3,4,5</sup>, Bettina Nadorp<sup>10,11</sup>, Benjamin Kroger<sup>12,13</sup>, Feng Hu<sup>14</sup>, Lewis Silverman<sup>2,4,5</sup>, Stephen S. Chung<sup>12,15</sup>, Elvin Wagenblast<sup>1,2,3,5</sup>, Ronan Chaligne<sup>16,17</sup>, Ann-Kathrin Eisfeld<sup>18</sup>, Deniz Demircioglu<sup>1,2,5,6</sup>, Dan A. Landau<sup>16,17</sup>, Piro Lito<sup>14</sup>, Elli Papaemmanuil<sup>7</sup>, Courtney D. DiNardo<sup>9</sup>, Dan Hasson<sup>1,2,3,5,6</sup>, Marina Konopleva<sup>19,20,21</sup> & Eirini P. Papapetrou<sup>1,2,3,4,5✉</sup>

Nature | Vol 636 | 5 December 2024 |

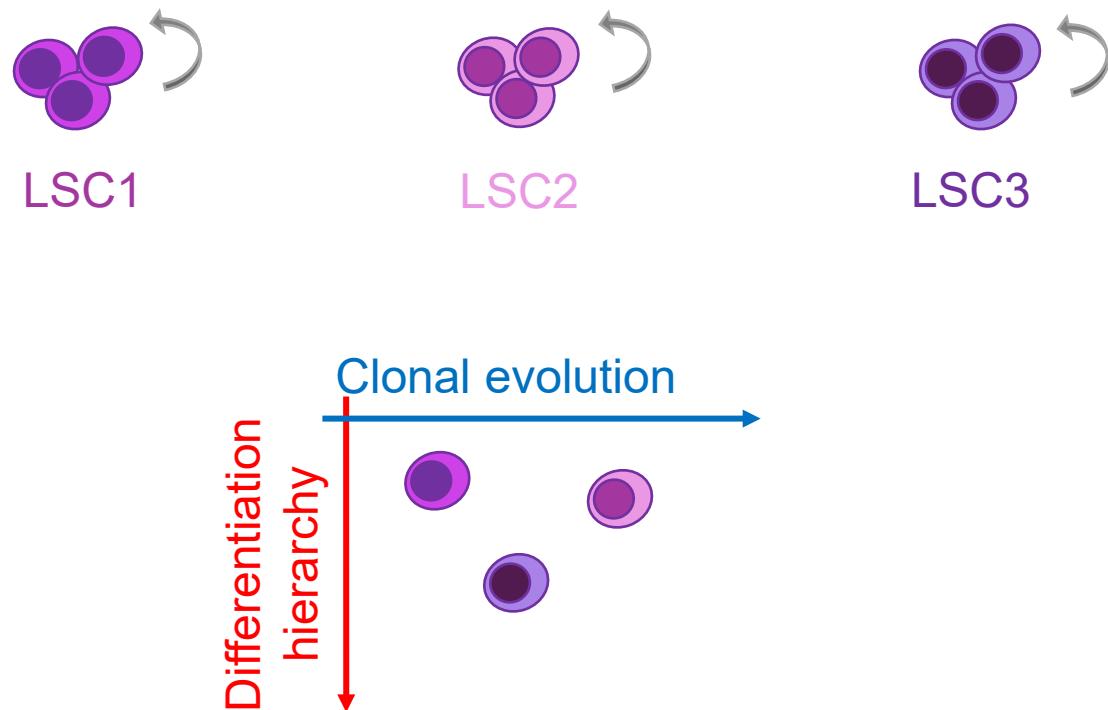
# LSCs in two dimensions



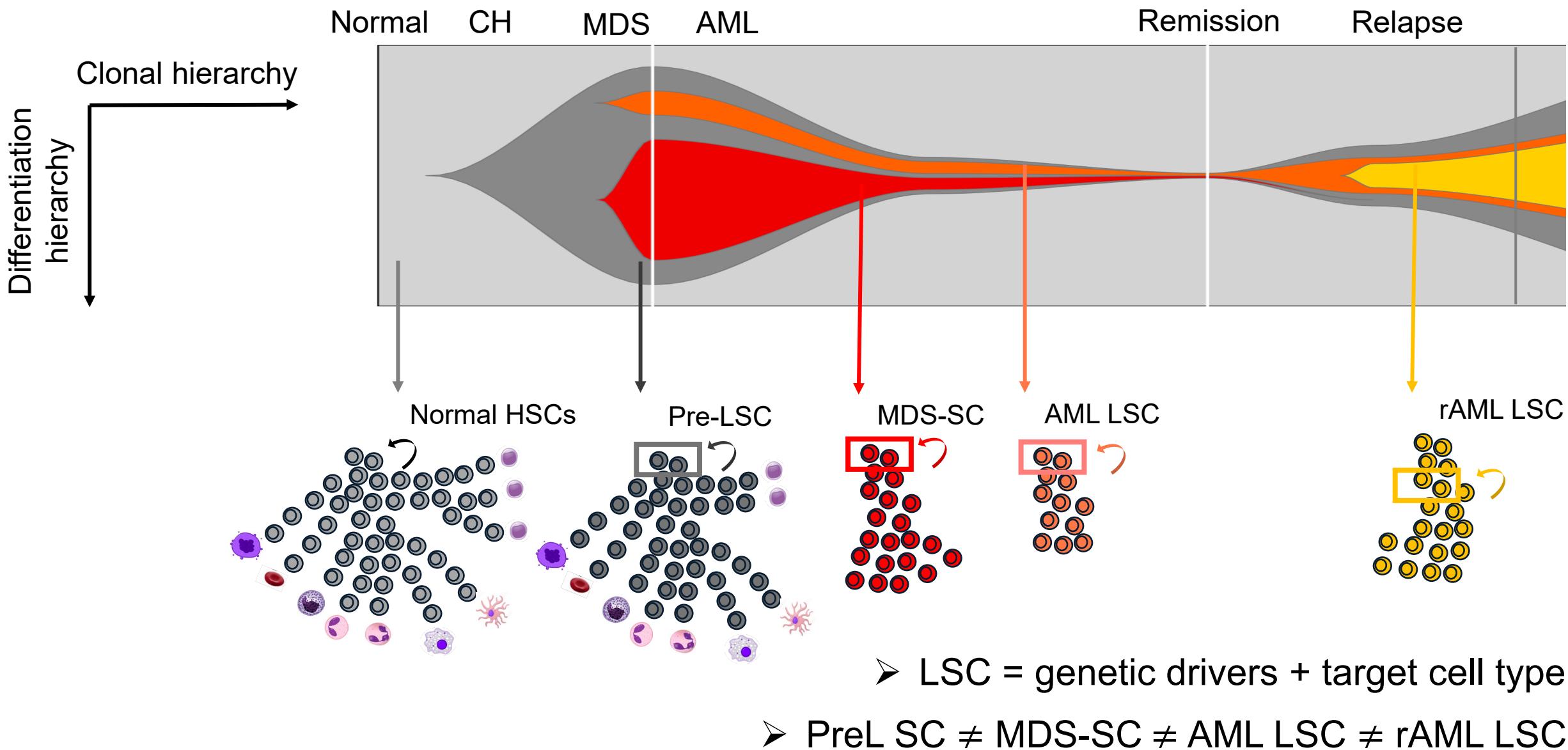
# LSCs in two dimensions



# LSCs in two dimensions



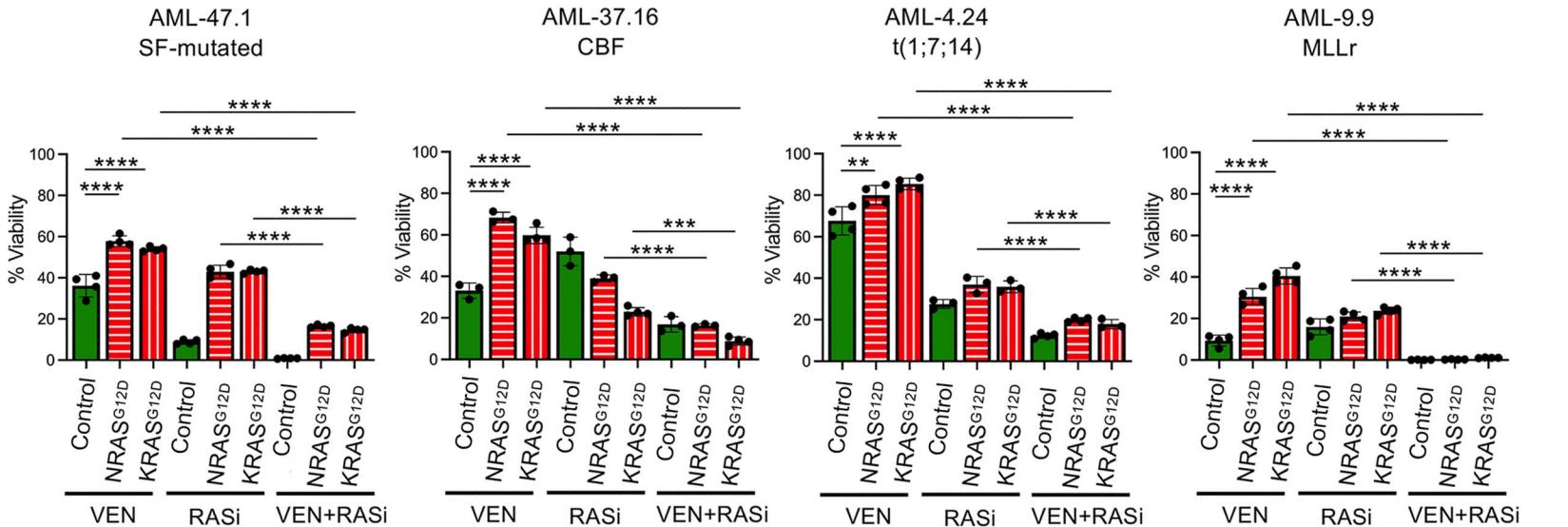
# A revised LSC model



- How can we overcome VEN resistance in RAS-MT AML?

# N/KRAS mutations confer VEN resistance to AML LSCs

RMC-7977  
active state-  
selective RAS  
multi-inhibitor

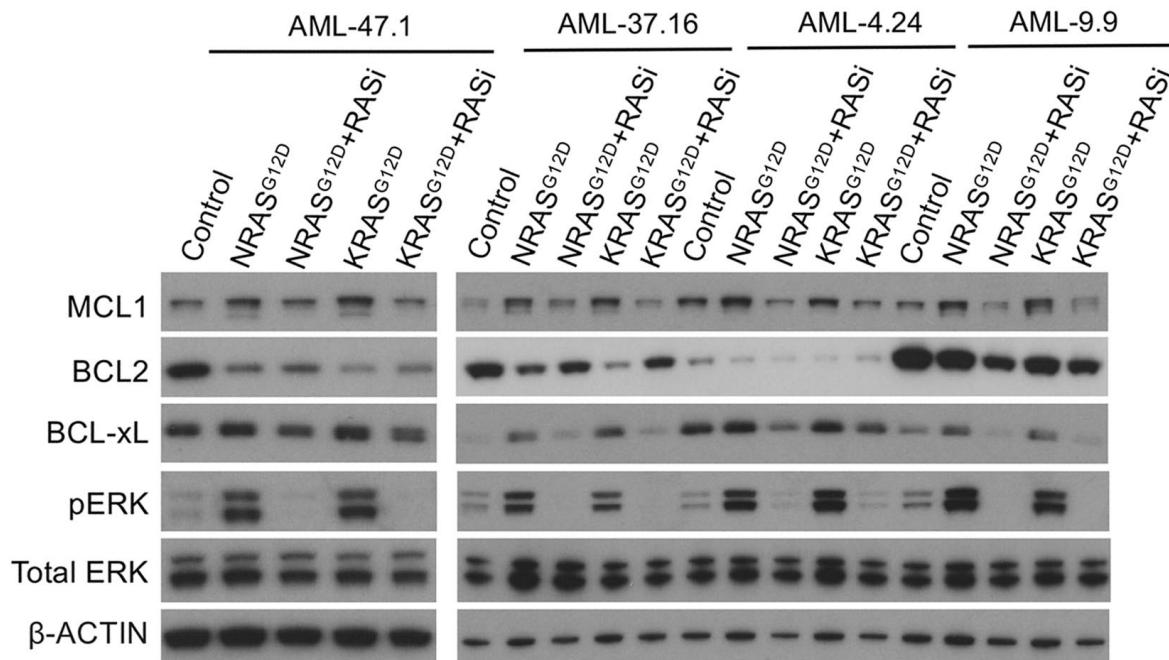


## Article

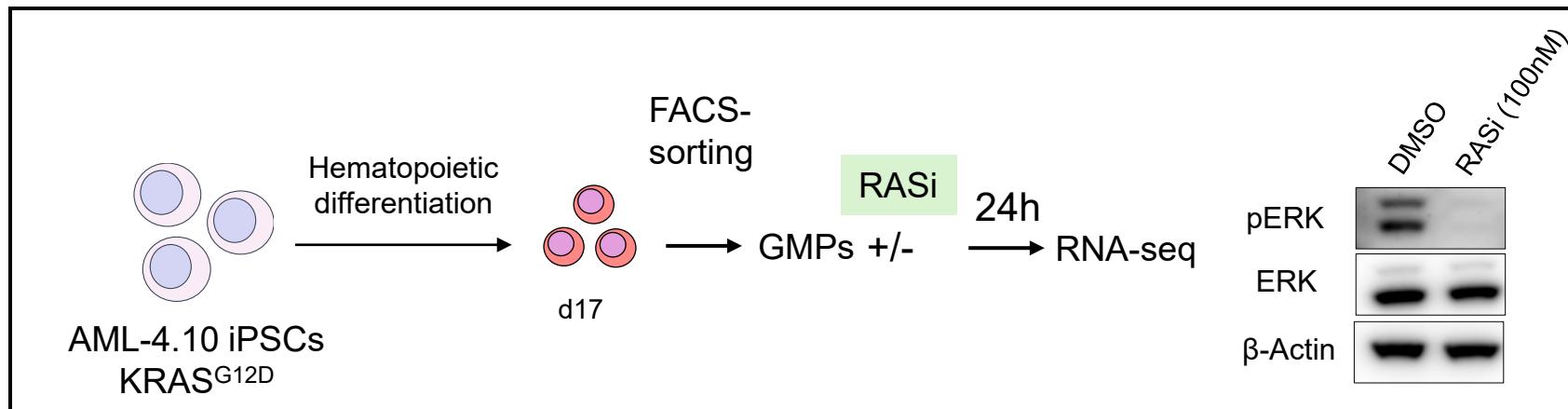
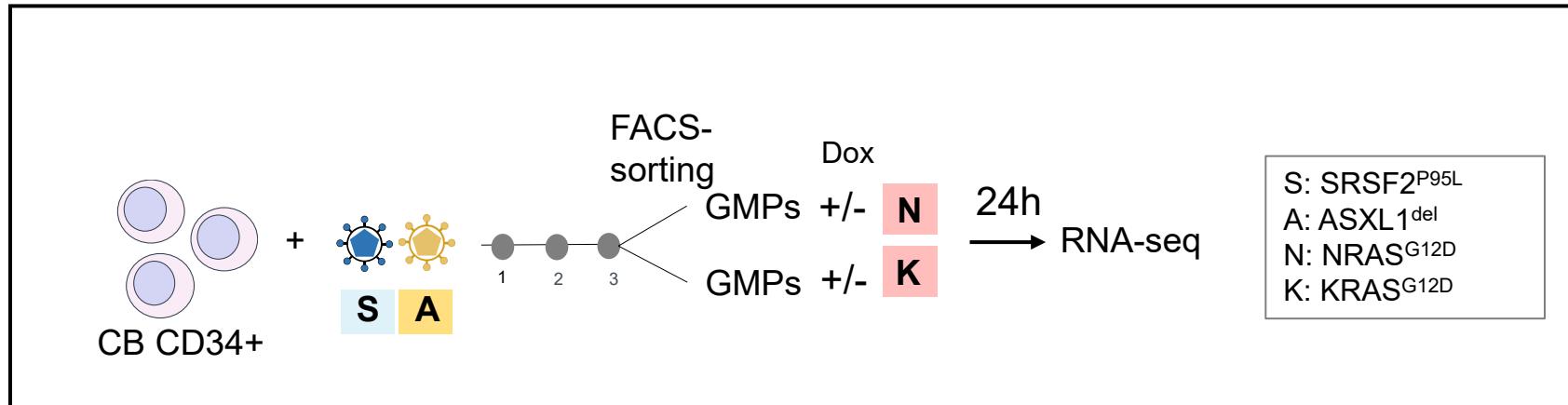
### Concurrent inhibition of oncogenic and wild-type RAS-GTP for cancer therapy

Matthew Holderfield<sup>1</sup>, Bianca J. Lee<sup>1</sup>, Jingjing Jiang<sup>1</sup>, Aidan Tomlinson<sup>1</sup>, Kyle J. Seamon<sup>1</sup>, Alessia Mira<sup>2</sup>, Enrico Patrucco<sup>2</sup>, Grace Goodhart<sup>3</sup>, Julien Dilly<sup>4</sup>, Yevgeniy Gindin<sup>1</sup>, Nuntana Dinglasan<sup>1</sup>, Yingyun Wang<sup>1</sup>, Lick Pui Lai<sup>1</sup>, Shurui Cai<sup>1</sup>, Lingyan Jiang<sup>1</sup>, Nicole Nasholm<sup>1</sup>, Nataliya Shifrin<sup>1</sup>, Cristina Blaj<sup>1</sup>, Harshit Shah<sup>1</sup>, James W. Evans<sup>1</sup>, Nilufar Montazer<sup>1</sup>, Oliver Lai<sup>1</sup>, Jade Shi<sup>1</sup>, Ethan Ahler<sup>1</sup>, Elsa Quintana<sup>1</sup>, Stephanie Chang<sup>1</sup>, Anthony Salvador<sup>1</sup>, Abby Marquez<sup>1</sup>, Jim Cregg<sup>1</sup>, Yang Liu<sup>1</sup>, Anthony Milin<sup>1</sup>, Anqi Chen<sup>1</sup>, Tamar Bar Ziv<sup>1</sup>, Dylan Parsons<sup>1</sup>, John E. Knox<sup>1</sup>, Jennifer E. Klomp<sup>5</sup>, Jennifer Roth<sup>6</sup>, Matthew Rees<sup>6</sup>, Melissa Ronan<sup>6</sup>, Antonio Cuevas-Navarro<sup>7</sup>, Feng Hu<sup>7</sup>, Piro Lito<sup>7,8</sup>, David Santamaria<sup>9</sup>, Andrew J. Aguirre<sup>4,6,10,11</sup>, Andrew M. Waters<sup>3,5,12,13</sup>, Channing J. Der<sup>5,12</sup>, Chiara Ambrogio<sup>2</sup>, Zhengping Wang<sup>1</sup>, Adrian L. Gill<sup>1</sup>, Elena S. Koltun<sup>1</sup>, Jacqueline A. M. Smith<sup>1</sup>✉, David Wildes<sup>1</sup>✉ & Mallika Singh<sup>1</sup>✉

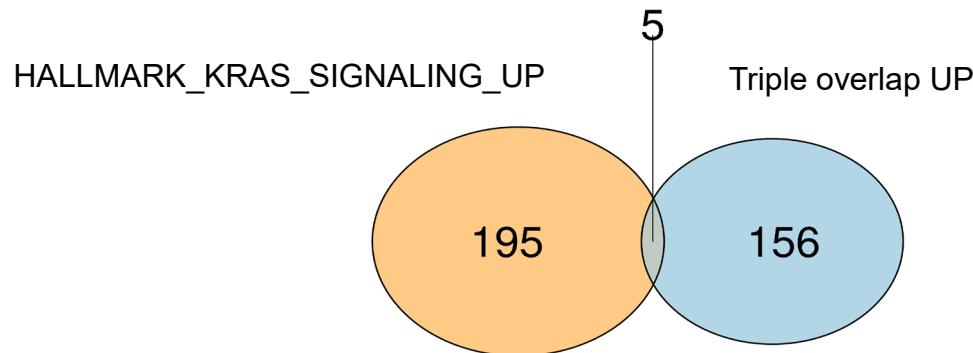
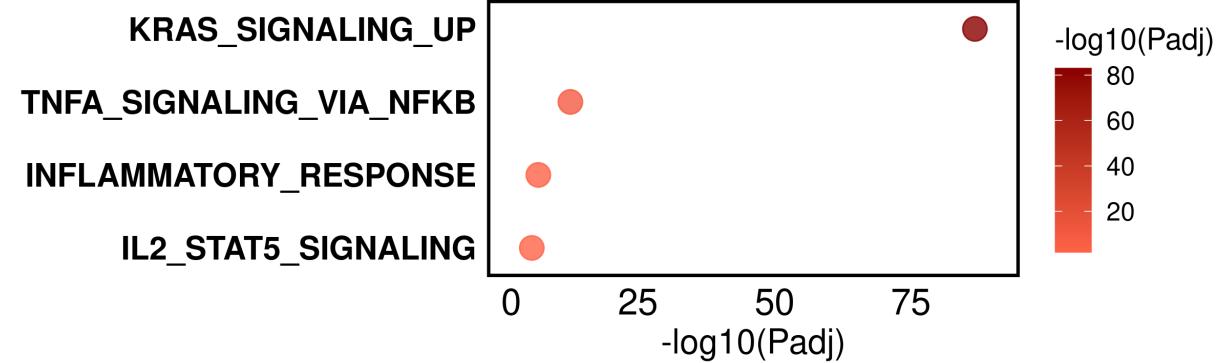
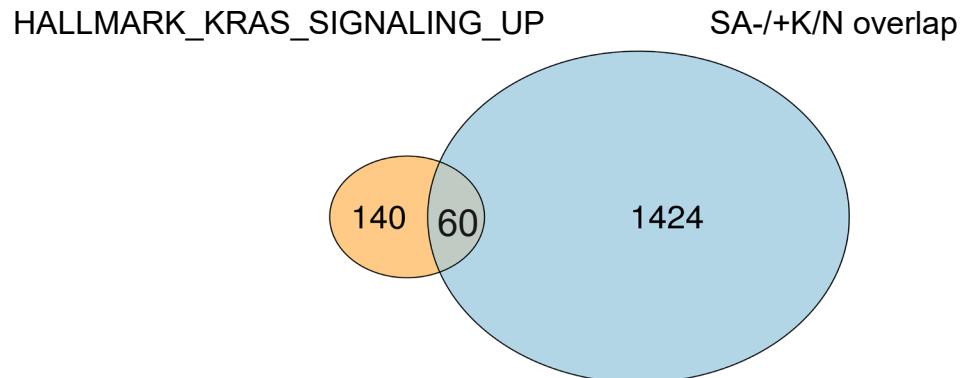
Nature | Vol 629 | 23 May 2024 |



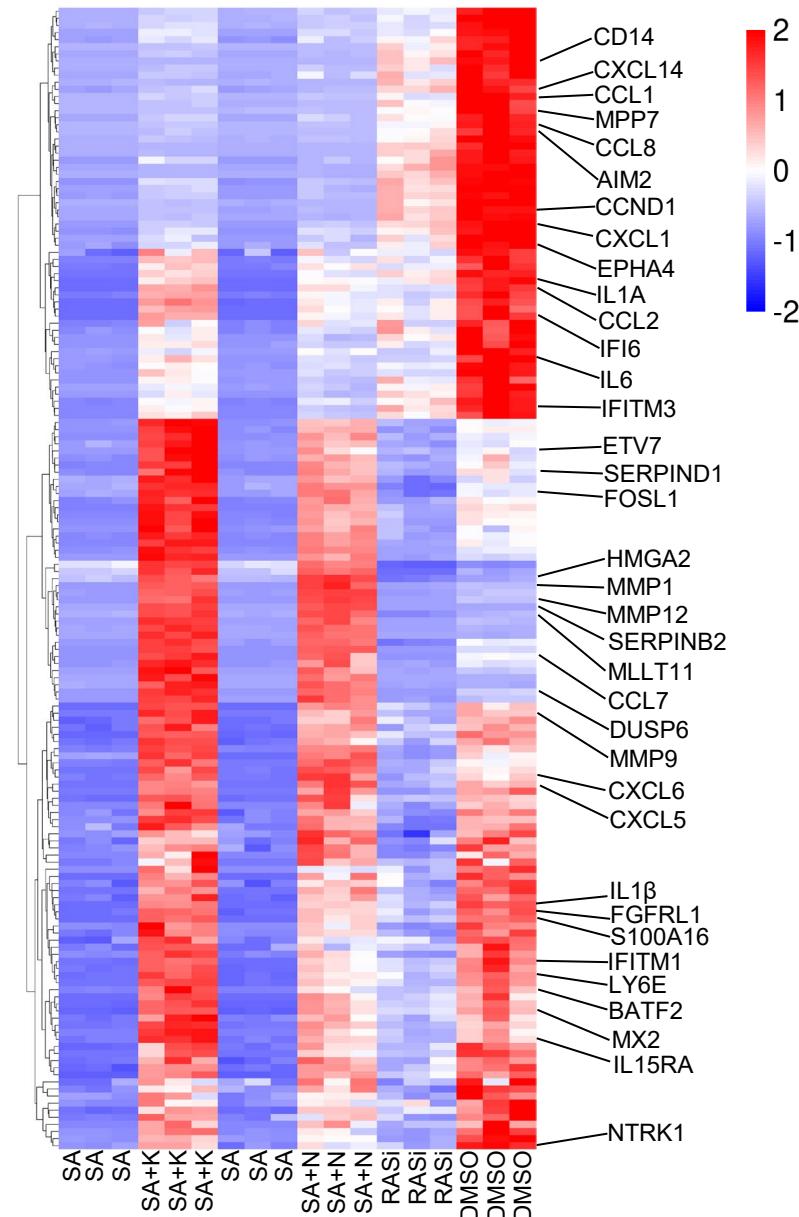
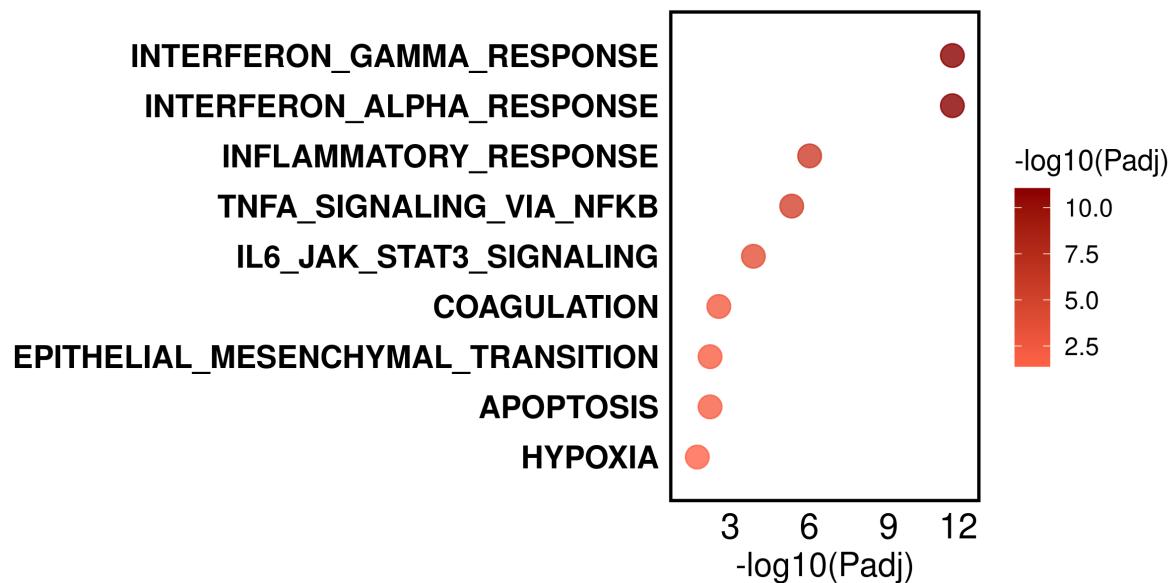
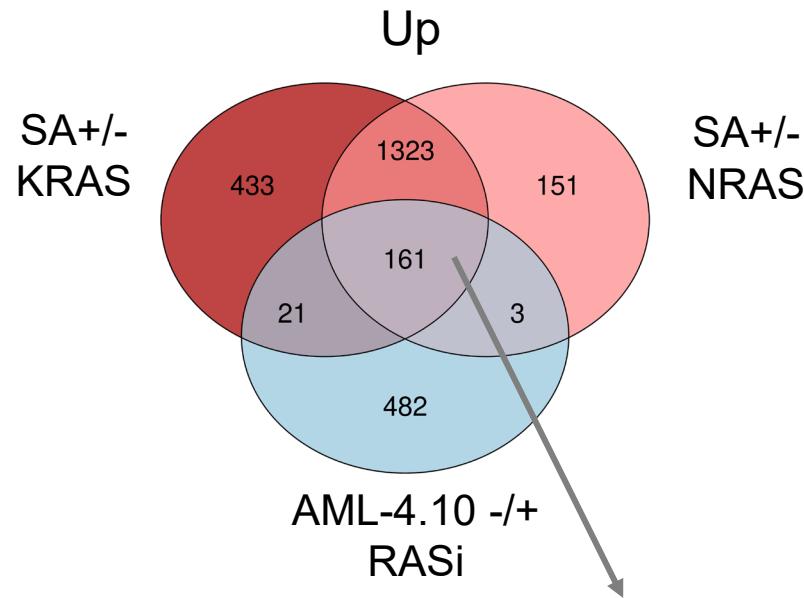
# A GMP LSC-specific RAS activation gene signature



# A GMP LSC-specific RAS activation gene signature



# A GMP LSC-specific RAS activation gene signature



**leukemia**  
**Papapetrou Lab**  
 differentiation  
**stem cells**  
 human myeloid cancer  
 iPSCs CRISPR  
 gene editing reprogramming  
 Cas9 CRISPR  
 RNA-Seq ATAC-Seq  
 single cell HSC ATAC-Seq  
 genomics CHIP  
 clonal AML  
 iPSCs CRISPR  
 gene editing mutation  
 reprogramming



NATIONAL  
CANCER  
INSTITUTE

LEUKEMIA &  
LYMPHOMA  
SOCIETY®  
fighting blood cancers

EvansMDS  
A funding initiative of  
The Edward P. Evans Foundation

Mount  
Sinai

The Tisch Cancer Institute



**BiNGS**  
Bioinformatics for Next  
Generation Sequencing

GABRIELLE'S  
ANGEL FOUNDATION  
FOR CANCER RESEARCH



Center for Advancement  
of **Blood Cancer Therapies**



### Papapetrou lab:

- Junya Sango, PhD
- Małgorzata Olszewska
- Hager Mansour, PhD
- Emmanuel Olivier, PhD
- Minh Nguyen, PhD
- Ben Jia, PhD
- Yujing Zhang, PhD
- Saul Carcamo, PhD
- Vrinda Jethalia
- Ivy Fan

### Papapetrou lab alumni:

- Diana Kotini, PhD
- Tiansu Wang, PhD
- Gulay Ulukaya, PhD
- Lewis Tomalin, PhD
- Tiansu Wang, PhD

### Collaborators:

- Marina Konopleva, MD PhD
- Abhi Maiti, MD
- Courtney DiNardo, MD
- Elli Papaemmanuil, PhD
- Dan Landau, PhD
- AK Eisfeld, MD
- Piro Lito, MD, PhD
- Elvin Wagenblast, PhD