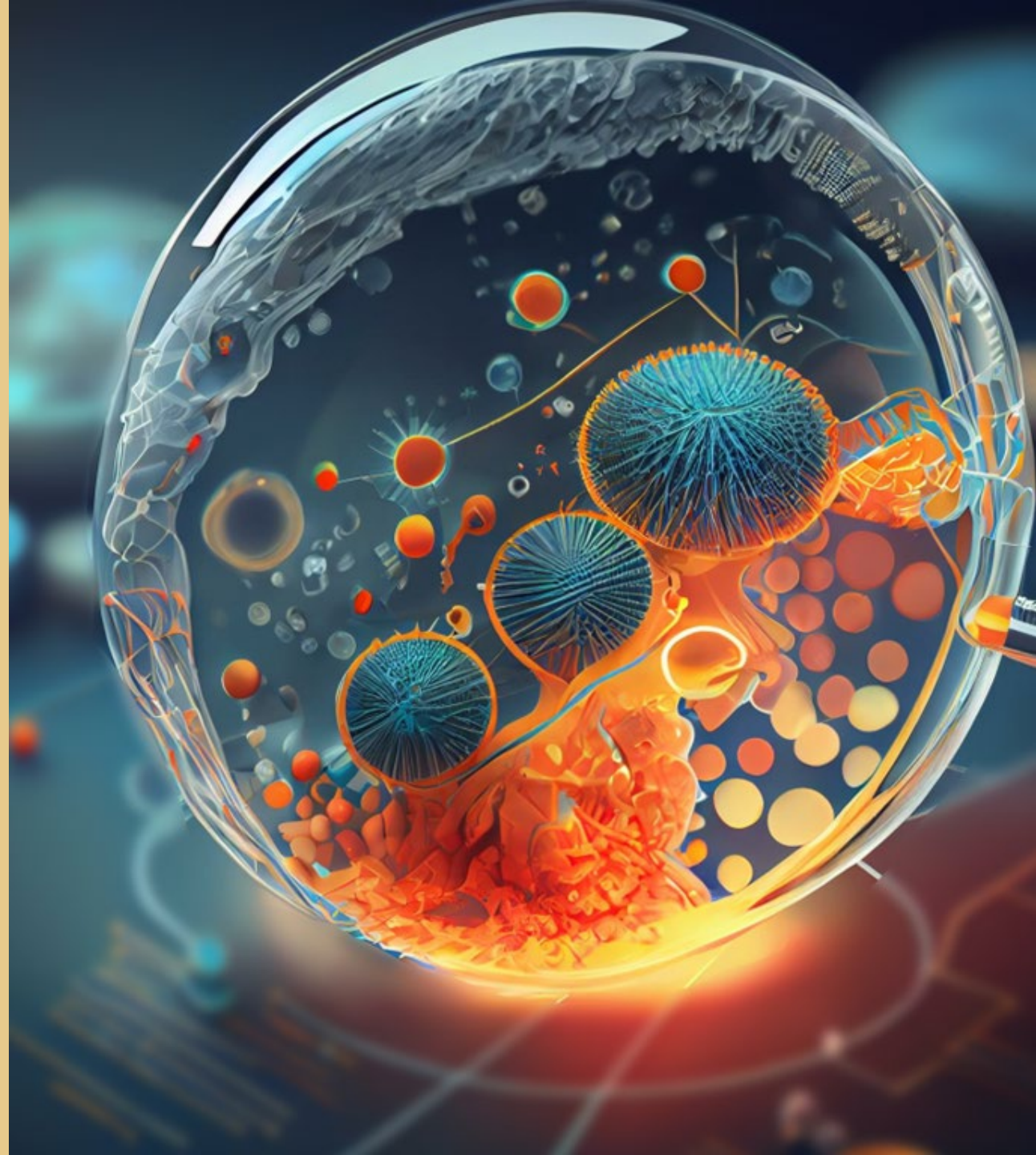


 eha **Sf(PM)**

From single-cell profiling to effective drug combinations

Applications to solid and
hematological cancers

September 26, 2024
Tero Aittokallio, PhD



Contents

- 01 Genomic + *f*PM platform
- 02 Current challenges
- 03 Clone-targeting treatments
- 04 Effective drug combinations
- 05 Single-cell analyses
- 06 Pilot study in AML (n=4)
- 07 Larger AML cohort (n=12)
- 08 Application to ovarian cancer
- 09 Biomarker discovery challenge
- 10 Conclusions

Tero Aittokallio, PhD

Group Leader, Institute for Cancer Research, Oslo University Hospital

Professor, OCBE, Department of Biostatistics, University of Oslo

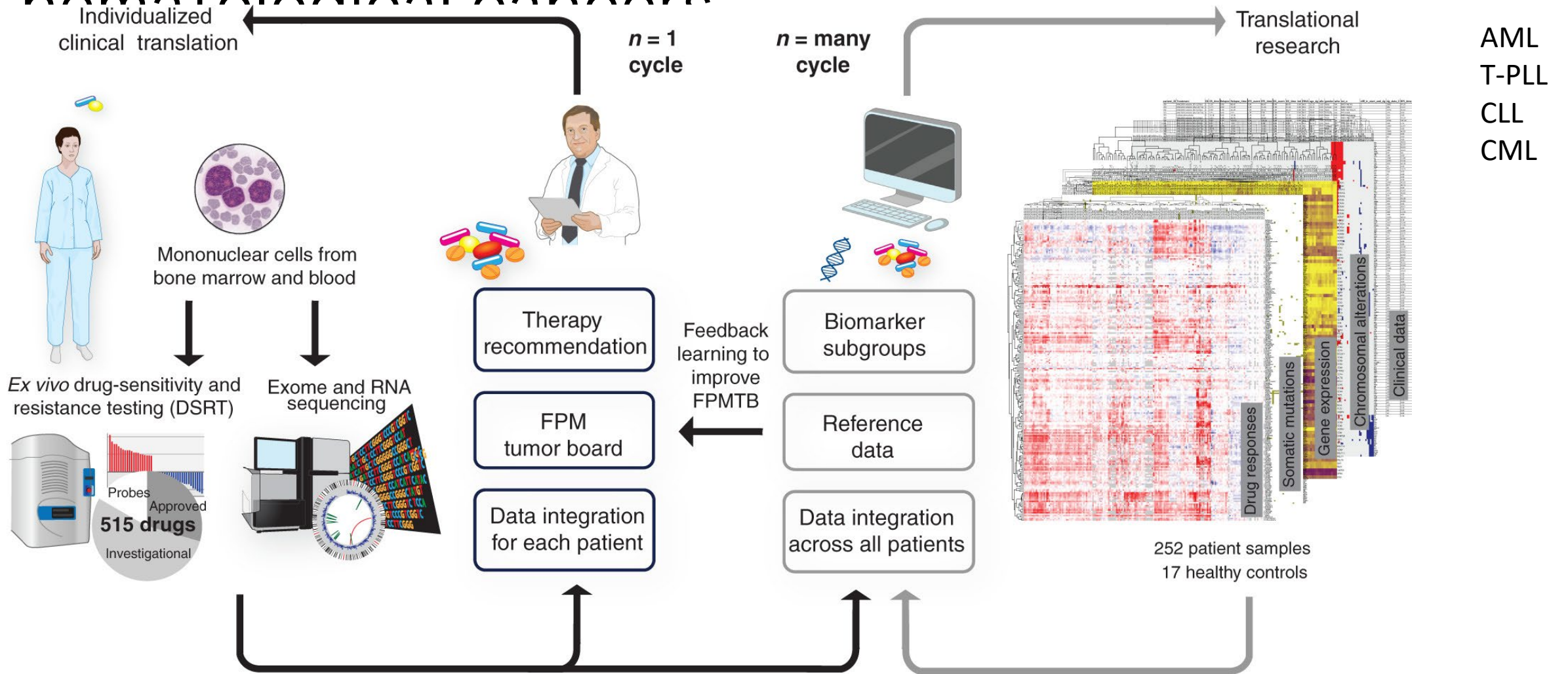
Group Leader, Institute for Molecular Medicine, University of Helsinki

Visiting Professor, InFLAMES Research Flagship, University of Turku

Disclosures: None



Functional precision medicine for hematological cancers



Current clinical and computational challenges

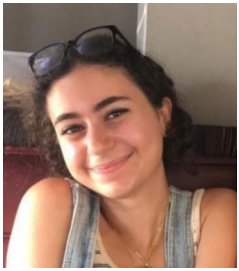
- 1) Modeling the effect of cellular heterogeneity of cancer sub-clones on clinical outcomes (e.g. treatment responses)
- 2) Identification of synergistic drug combinations that inhibit multiple cancer driving pathways and sub-populations
- 3) Mining of clinical, functional, and genomic biomarkers that are predictive of (combination) treatment responses

Response
predictive
biomarkers

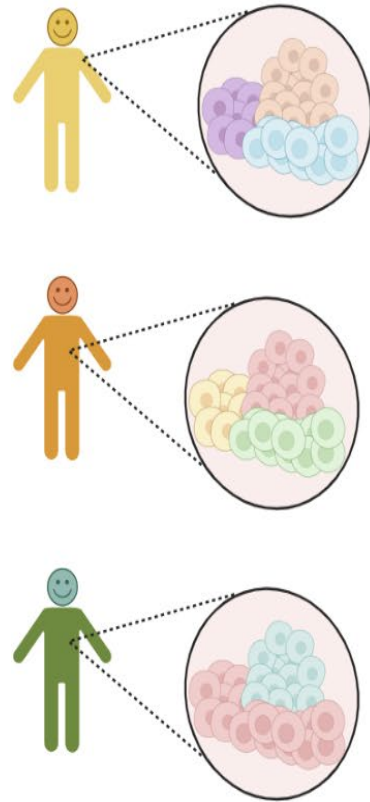


Optimal therapy
options for each
individual patient

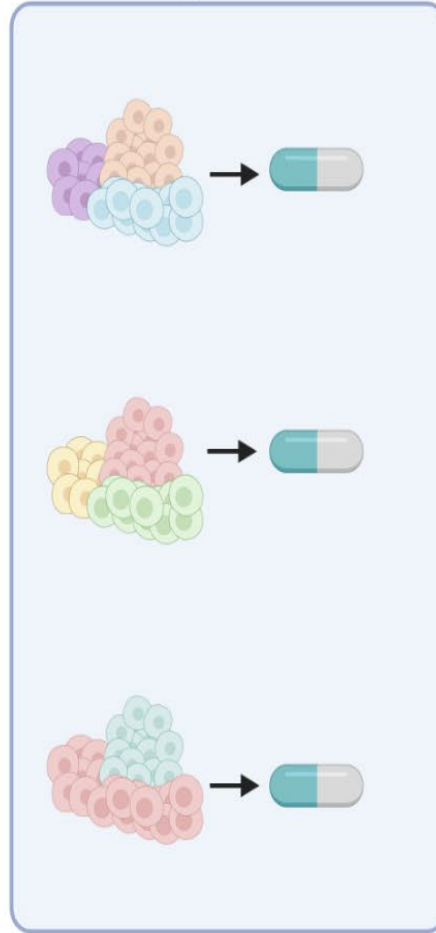
Targeted cancer therapy and tumour heterogeneity



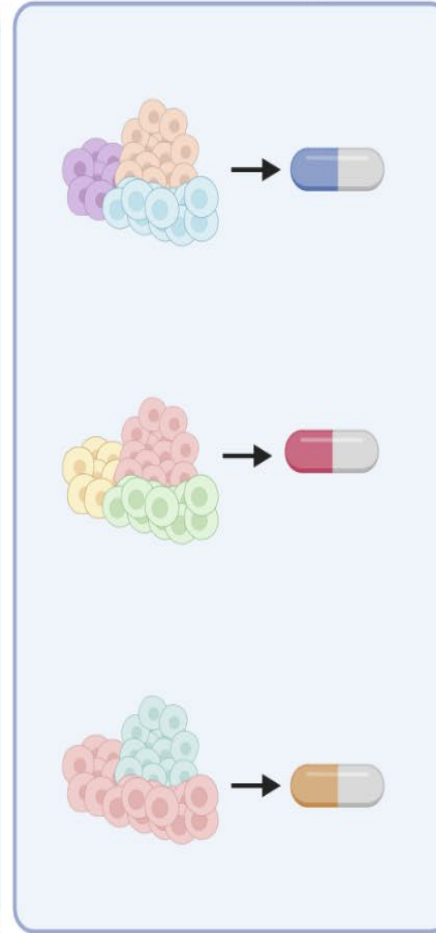
Kristen Nader



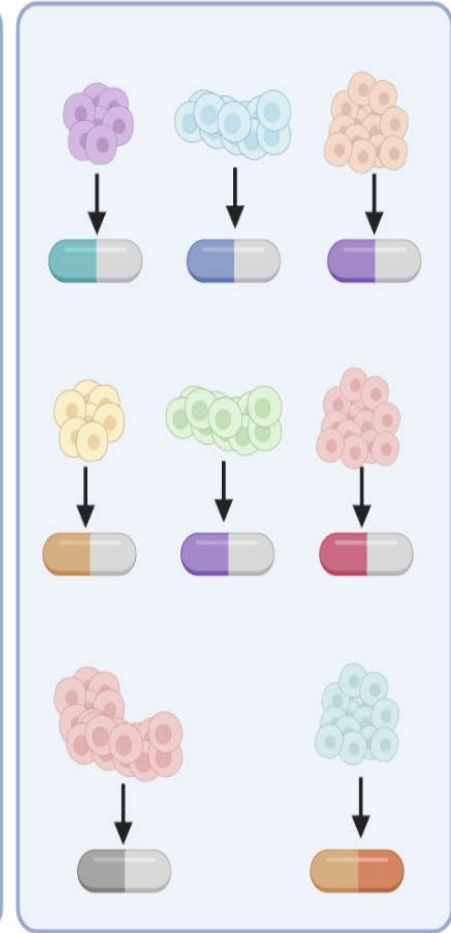
One Size Fits all approach



Targeted Monotherapy



Targeted Combination Therapy



AI-guided treatment selection?



Multi-modal modelling!



Credit: A2IDEA

How to identify successful drug combinations?

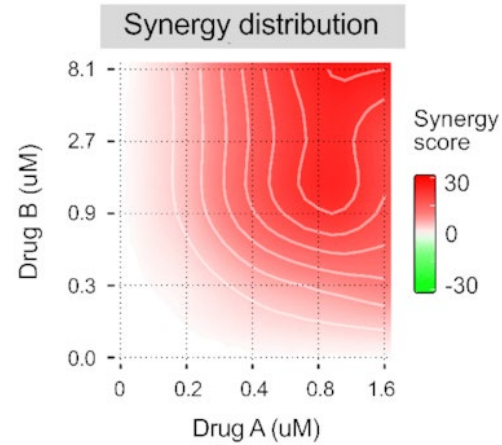


Anil Kumar Giri

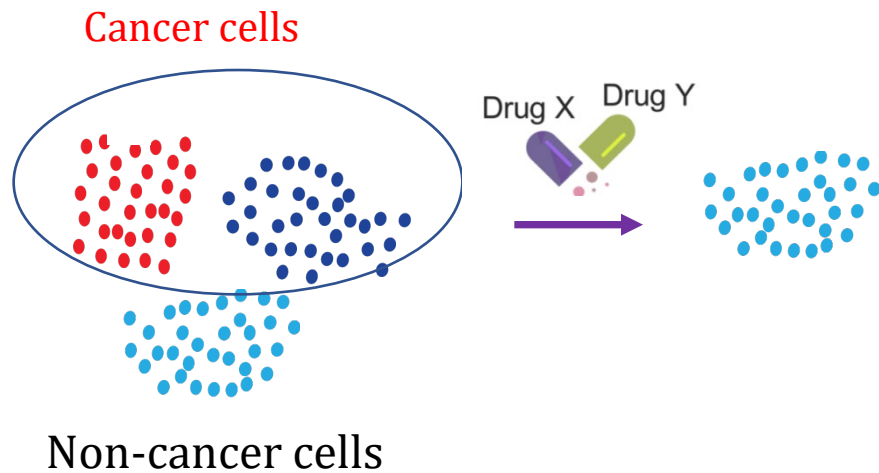
Effective



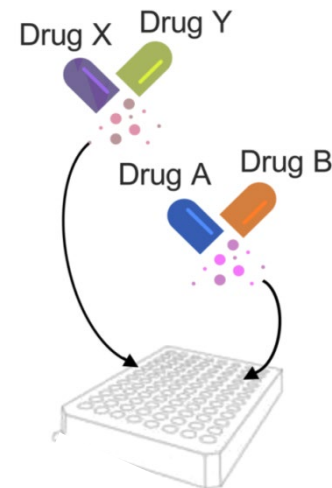
Synergistic



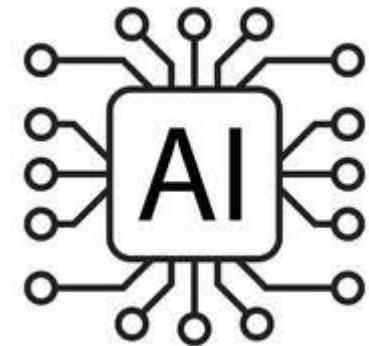
Cancer cell selective (less toxic)



Testing in patient-derived cells



Scarce primary cells



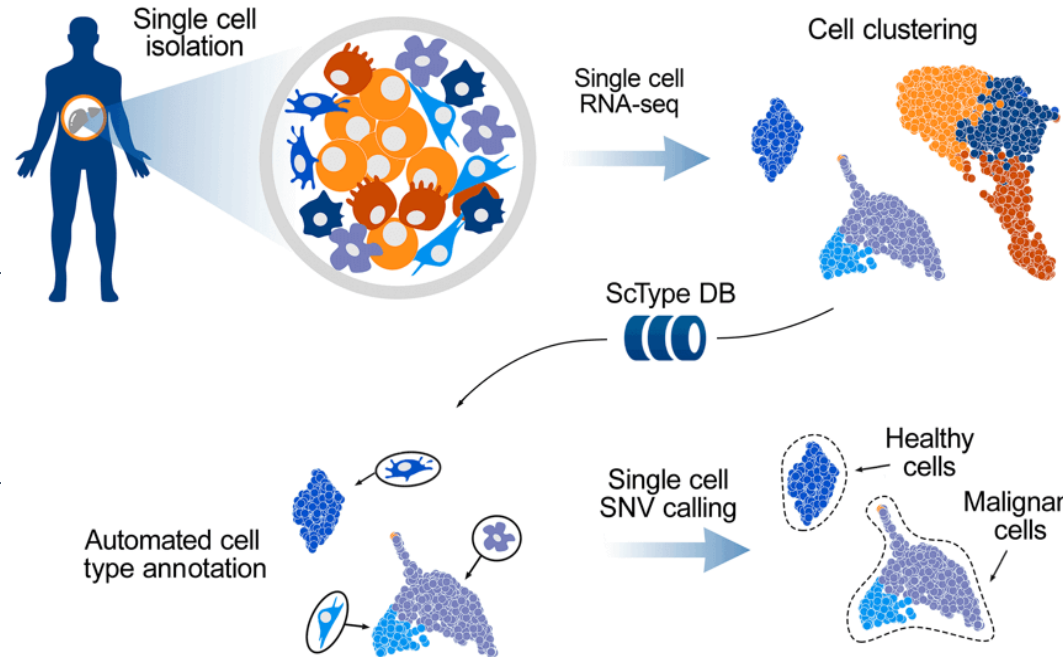
ScType web-app and marker database for fully-automated and ultra-fast cell type identification



Aleksandr Ianevski

Input data

scRNA-seq expression matrix



Output

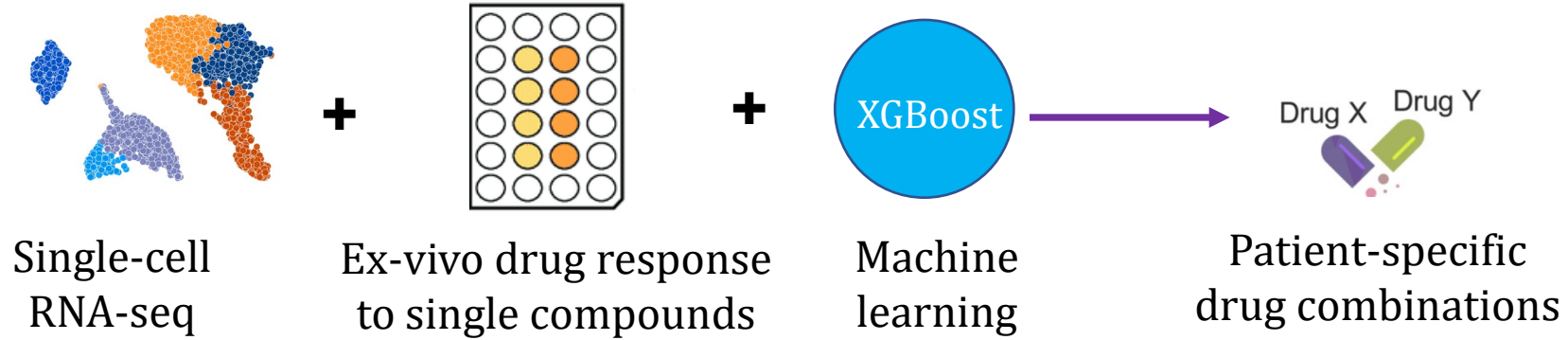
Automated cell type annotations

<https://sctype.app>

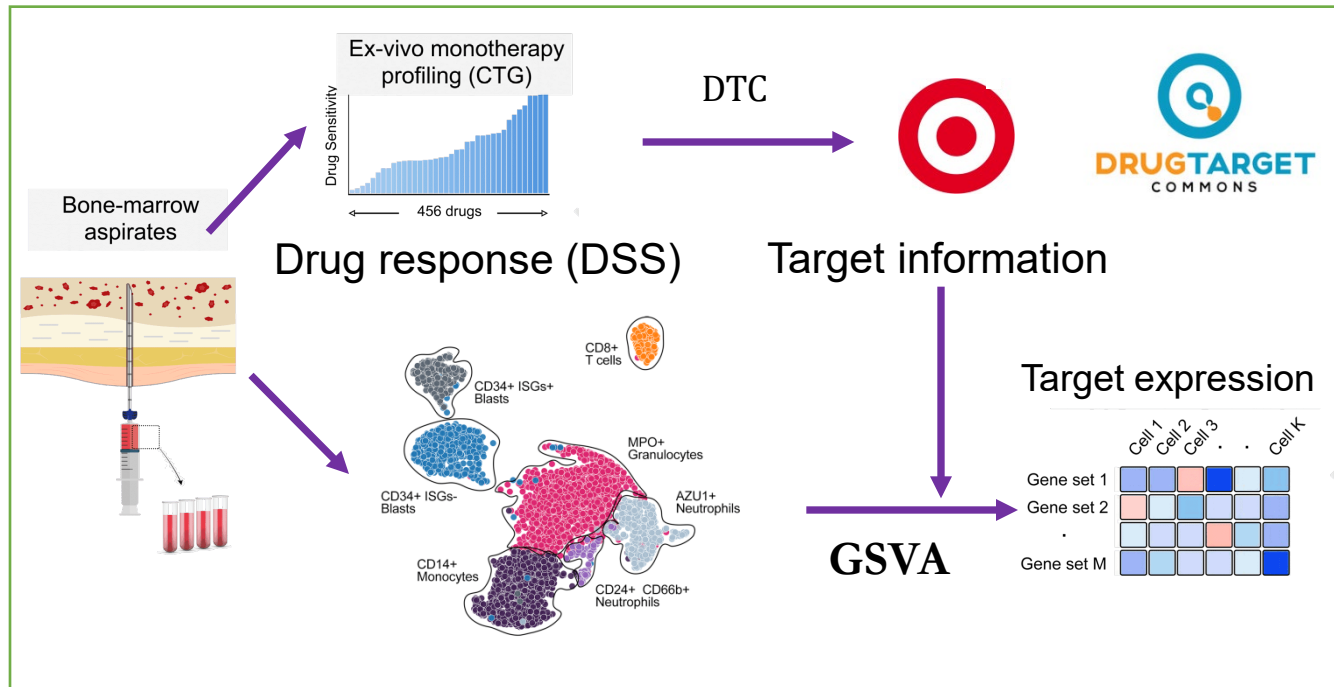
> 270 citations

Ianevski et al., Nat Comm 2022

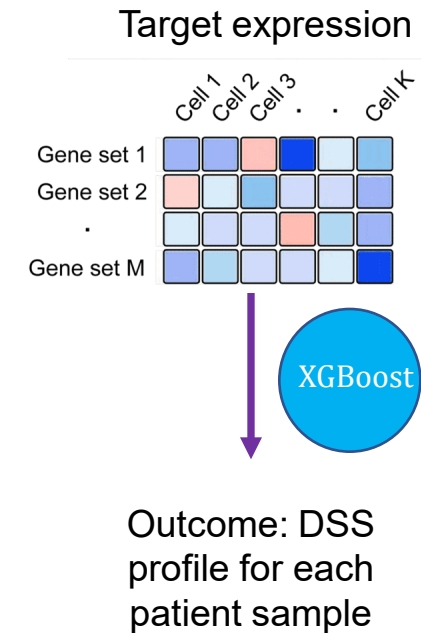
ML algorithm to tailor drug combinations for individual patients



Data processing

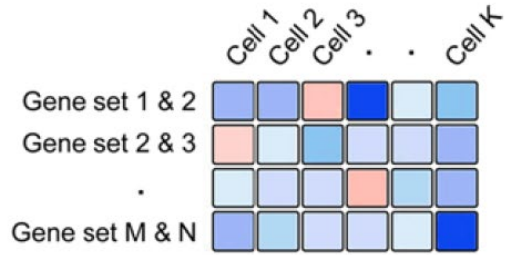


Machine learning

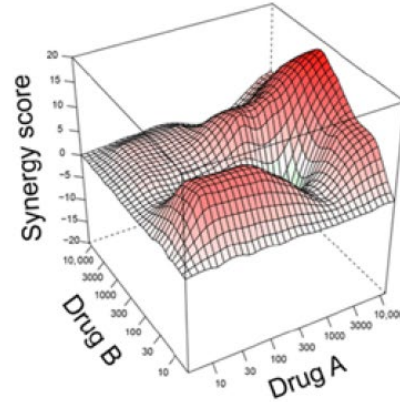


Patient-specific testing using *ex vivo* drug testing assays

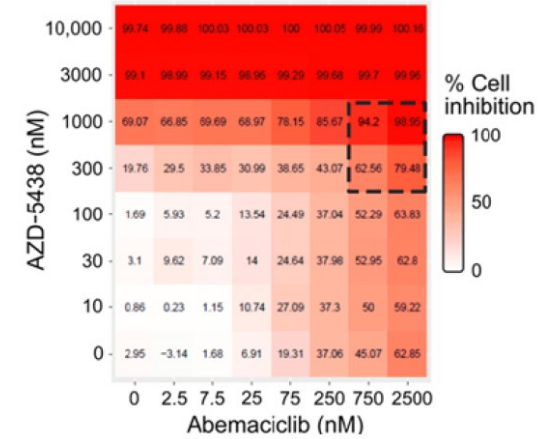
GSVA of drug combination target gene sets



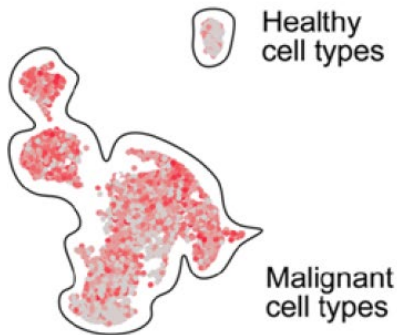
Prediction of synergistic, potent, and low-toxicity combinations



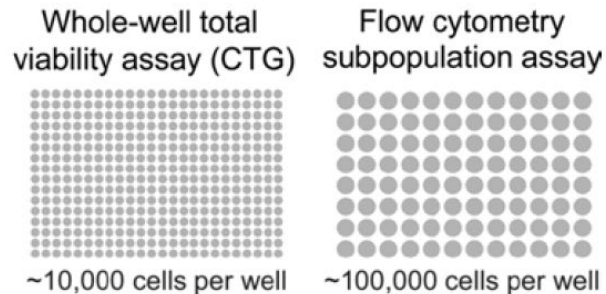
Dose-response matrix



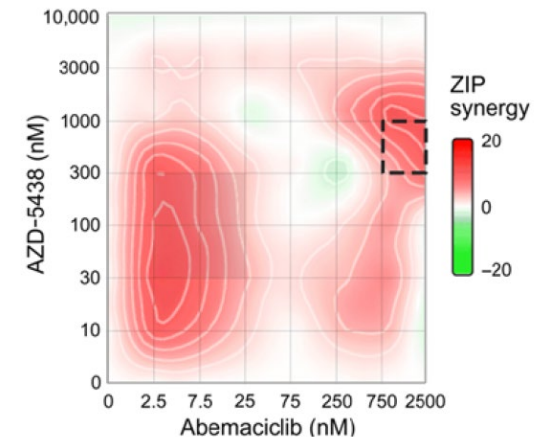
Combination target expression in malignant and healthy cells



Experimental validation of top predicted combinations



Synergy distribution



Prediction of patient-specific drug combinations in 4 longitudinal AML samples

Table 1. Clinical and molecular characteristics of the patients with AML and samples. Flow cytometry–based clinical immunophenotyping was performed in fresh red blood cell–depleted bone marrow samples at Helsinki University Hospital, whereas the scRNA-seq profiling was done on the basis of thawed Ficoll-treated and cryopreserved AML cells at the FIMM (Institute for Molecular Medicine Finland) Single-Cell Unit. The blast percentages differ between the two readouts because of different cell isolation protocols and different blast identification approaches. ICD-O, International Classification of Diseases for Oncology.

Patient sample	Disease stage	Age	Sex	scRNA-based blast (%)	Clinical morphological blast (%)	Diagnosis (ICD-O)	FAB type*	Previous malignancies or predisposing conditions	Risk class at the time of diagnosis	Potential driver mutations	Treatment history
AML1_D	Diagnosis	35	M	68	65	AML, C92, 9874	M2	No	Low	<i>WT1</i> , <i>CCND2</i> , and <i>CEBPA</i>	Cytarabine- idarubicin, lenalidomide
AML2_R	Refractory	68	M	26	40	AML C92, 9920	NA	Non-Hodgkin's lymphoma	Intermediate	<i>DNMT3A</i> , <i>ERG</i> , <i>U2AF1</i> , and <i>BCOR</i>	Cytarabine- idarubicin, azacytidine
AML3_D	Diagnosis	70	F	35	70	AML C92	M1	Non-Hodgkin's lymphoma	Intermediate	<i>NPM1</i> and <i>TET2</i>	Azacitidine
AML3_R	Refractory	71	F	56	42	AML, C92	M1	Non-Hodgkin's lymphoma	Intermediate	<i>NPM1</i> , <i>TET2</i> , and <i>HDAC 1,2,7</i>	Azacitidine- venetoclax

*FAB, the French-American-British classification.



CANCER

Patient-tailored design for selective co-inhibition of leukemic cell subpopulations

Aleksandr Ianevski^{1,2}, Jenni Lahtela¹, Komal K. Javarappa¹, Philipp Sergeev¹, Bishwa R. Ghimire¹, Prson Gautam¹, Markus Vähä-Koskela¹, Laura Turunen¹, Nora Linnavirta¹, Heikki Kuusanmäki^{1,3,4}, Mika Kontro⁴, Kimmo Porkka⁵, Caroline A. Heckman¹, Pirkko Mattila¹, Krister Wennerberg^{1,3*}, Anil K. Giri^{1*}, Tero Aittokallio^{1,2,6,7*}

Source codes available at

<https://github.com/ianeovskiAleksandr/scComb>

Current developments

- Selective combination predictions using scRNA-seq data alone
- Applications to solid tumors (high-grade serous carcinoma patients)
- Combination testing in more advanced assays (organoids, PDX)

FIMM

HUS

HRUH
HEMATOLOGY
RESEARCH UNIT
HELSINKI



iCAN

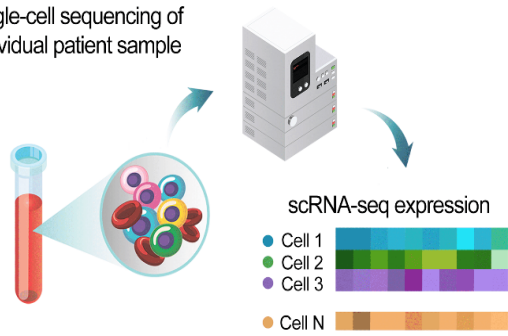
Patient-tailored therapies for selective co-inhibition of multiple cancer clones using single-cell transcriptomics alone

lanevski, Nader, et al. Nature Communications, to appear

<https://www.biorxiv.org/content/10.1101/2023.06.26.546571v1>

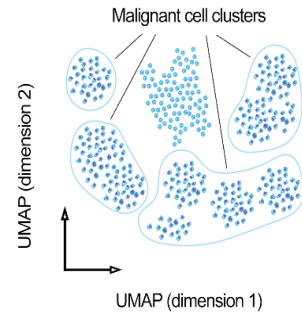
a

Single-cell sequencing of individual patient sample

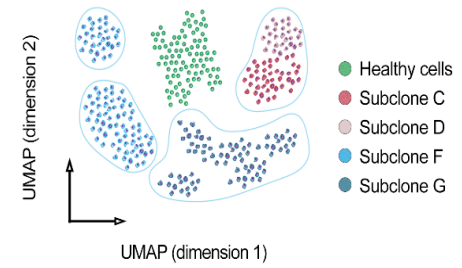


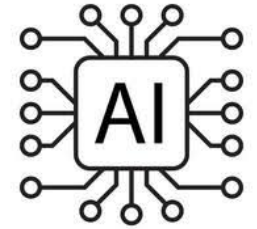
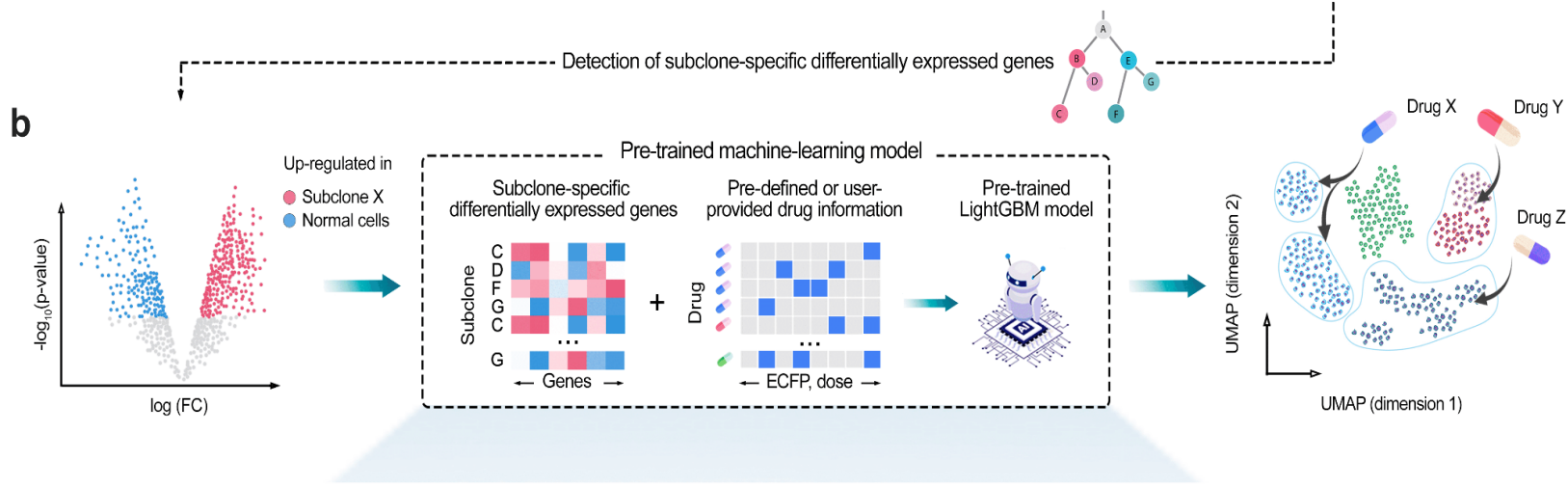
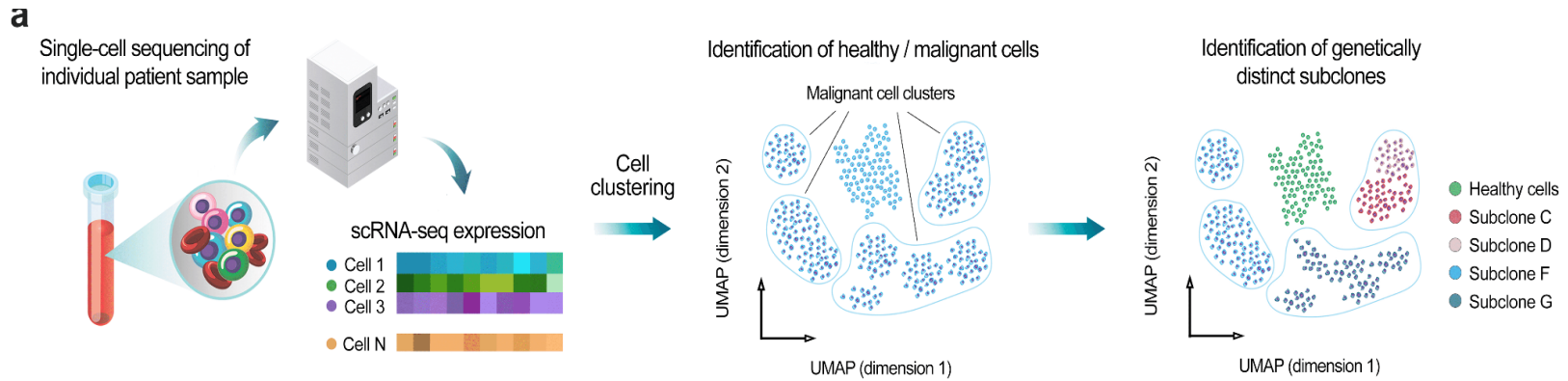
Cell clustering

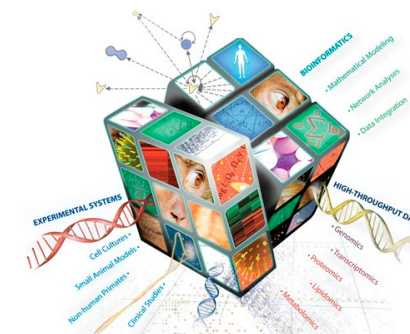
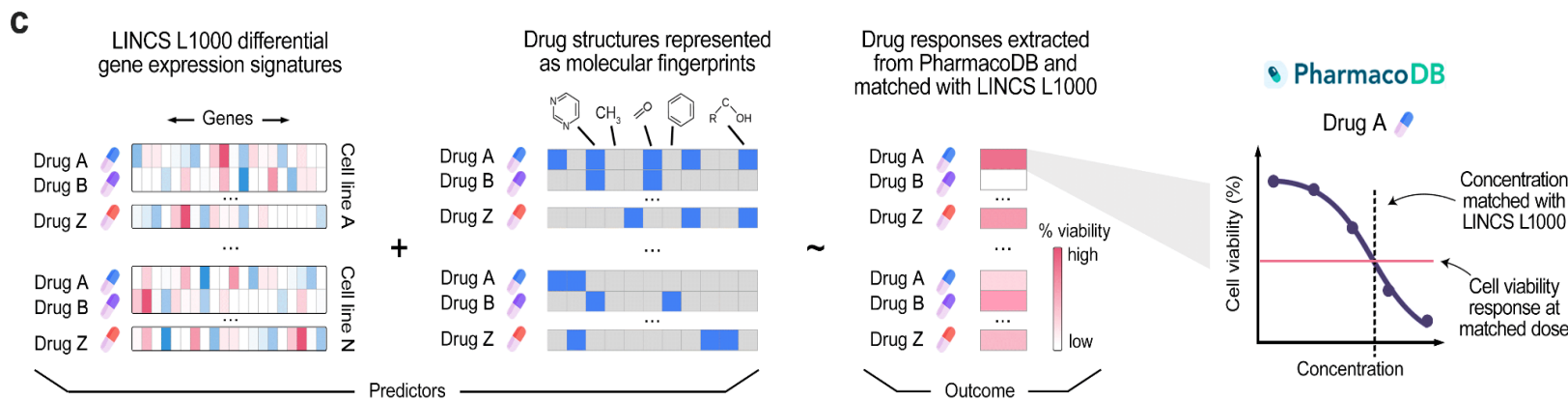
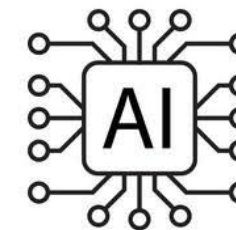
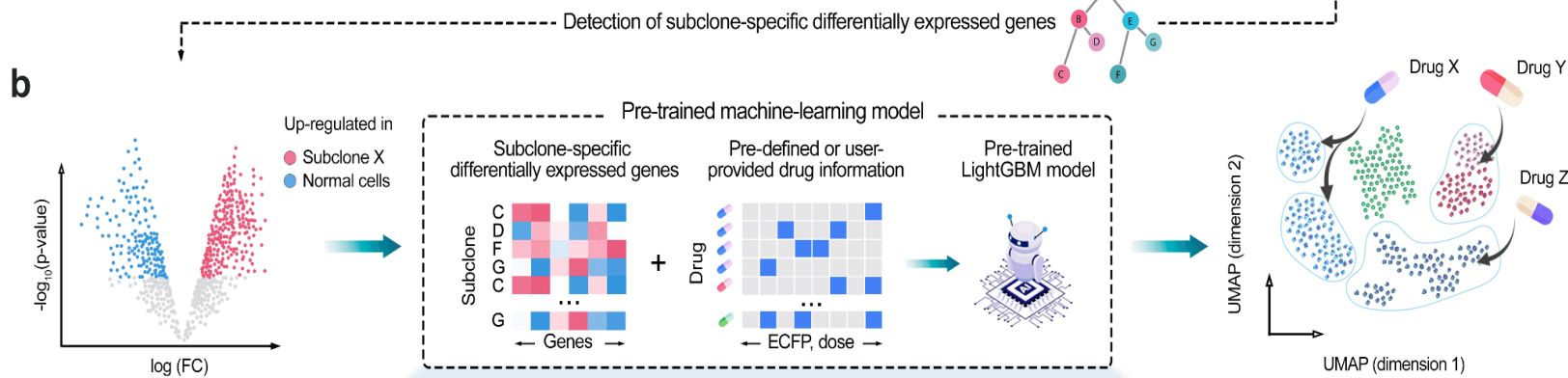
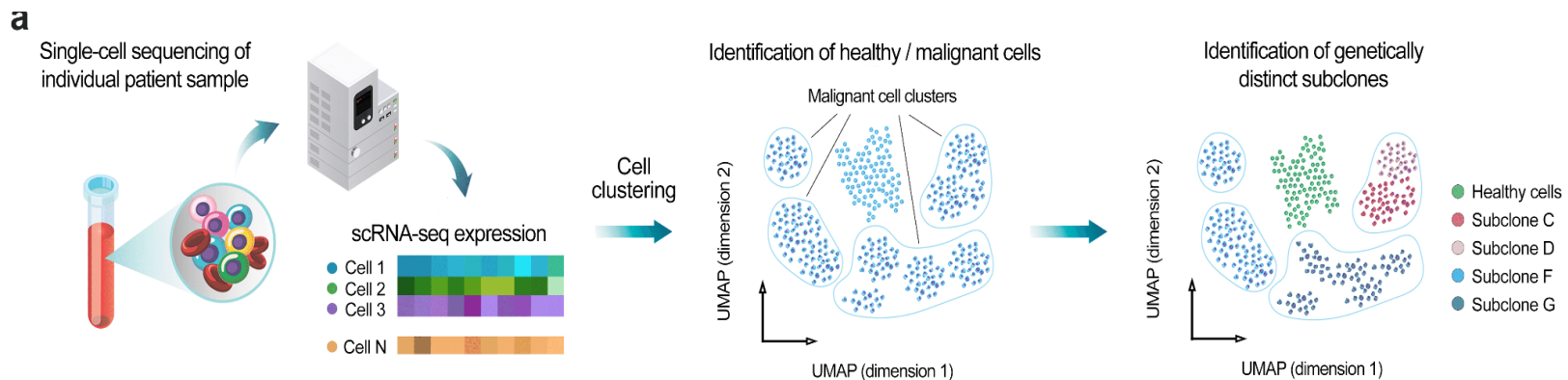
Identification of healthy / malignant cells



Identification of genetically distinct subclones





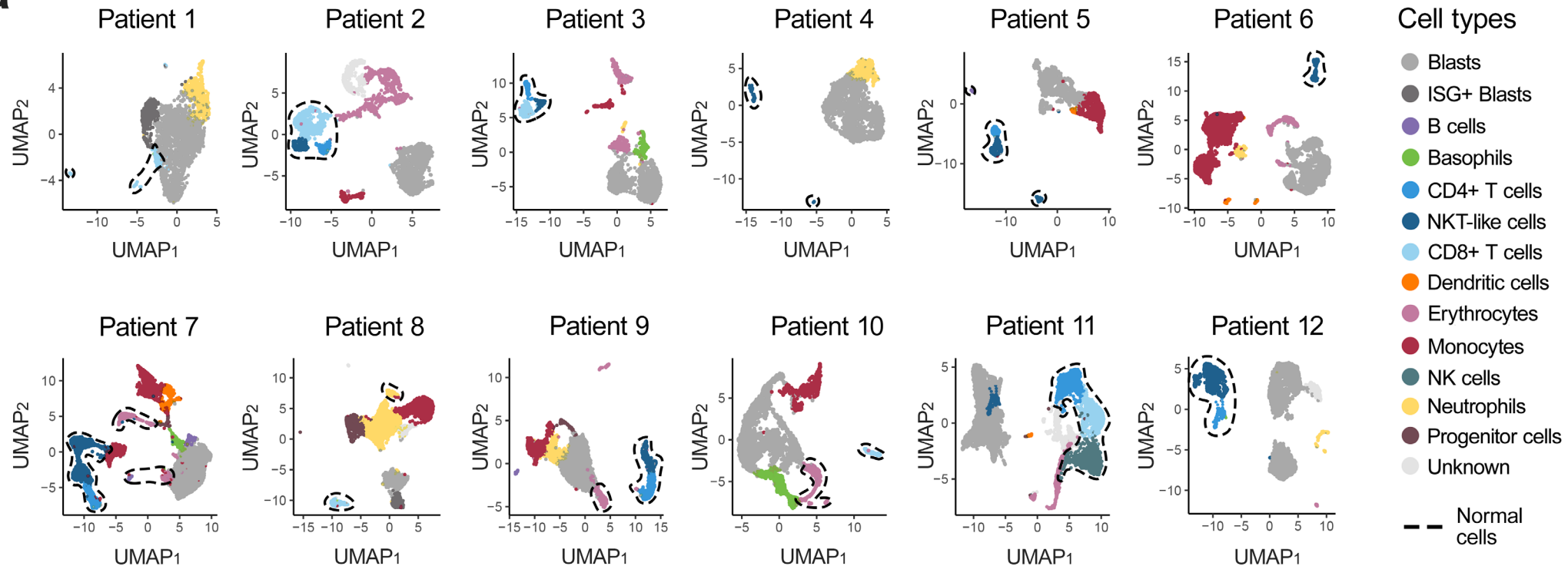


Heterogeneous cell type compositions across patients

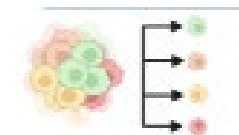
Need for personalized treatment options

Cell type annotation – ScType + CopyKAT + SCEVAN

a

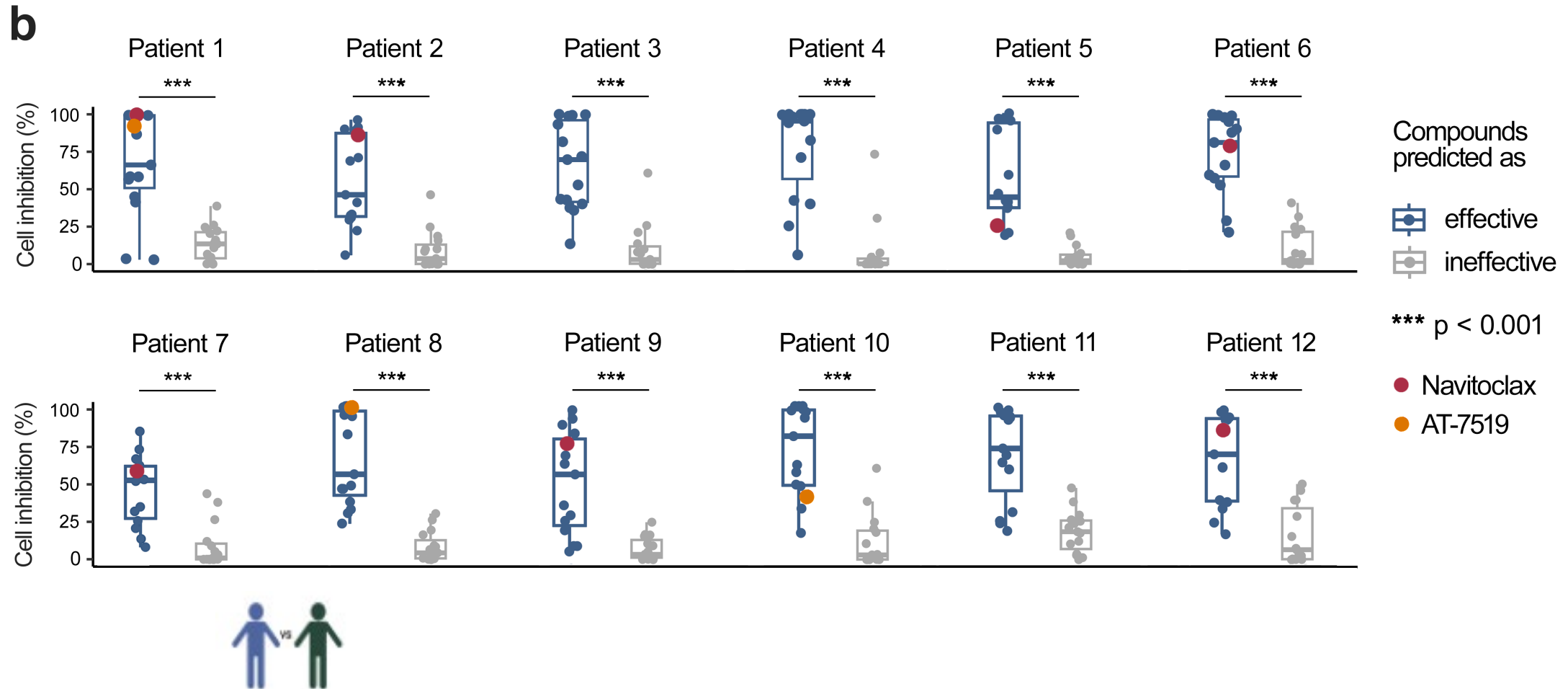


10x Genomics, FIMM single-cell core



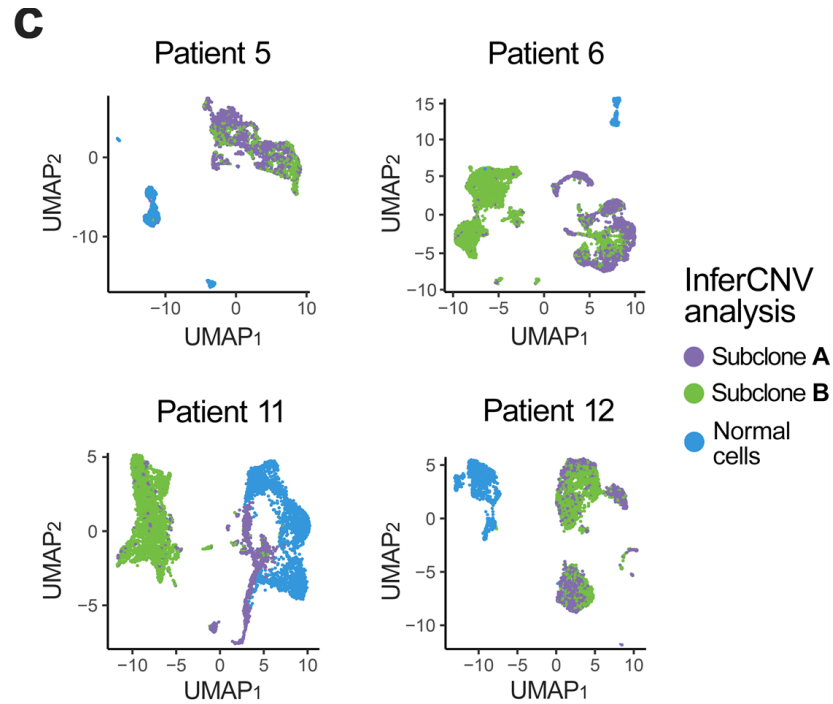
Retrospective validation:

Bulk CTG drug response profiling confirms that the model predicts effective single-agent treatments

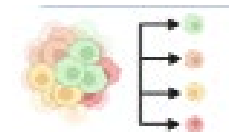
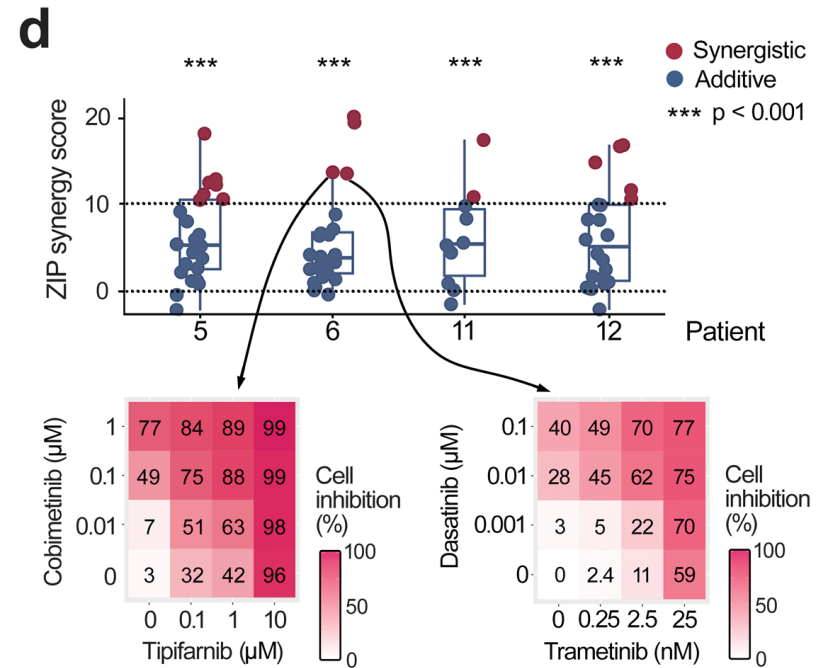


Prospective validation: Patient- and clone-specific drug combination predictions in four AML patients

Identify subclones - InferCNV



Testing patient-specific combinations



Flow cytometry cell population drug assay

Cancer cell-selective co-inhibition effects

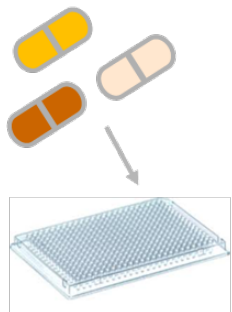


Tanja Ruokoranta

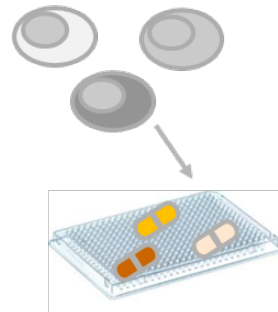


Heikki Kuusanmäki

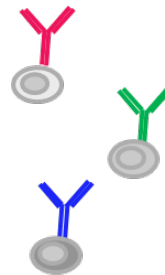
Drug administration



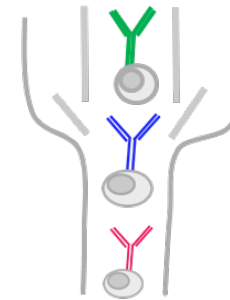
Cell dispensing



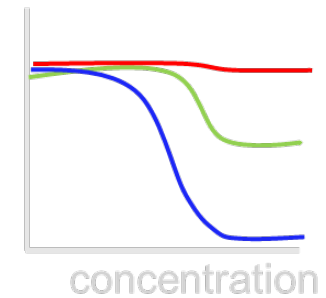
Staining



Readout

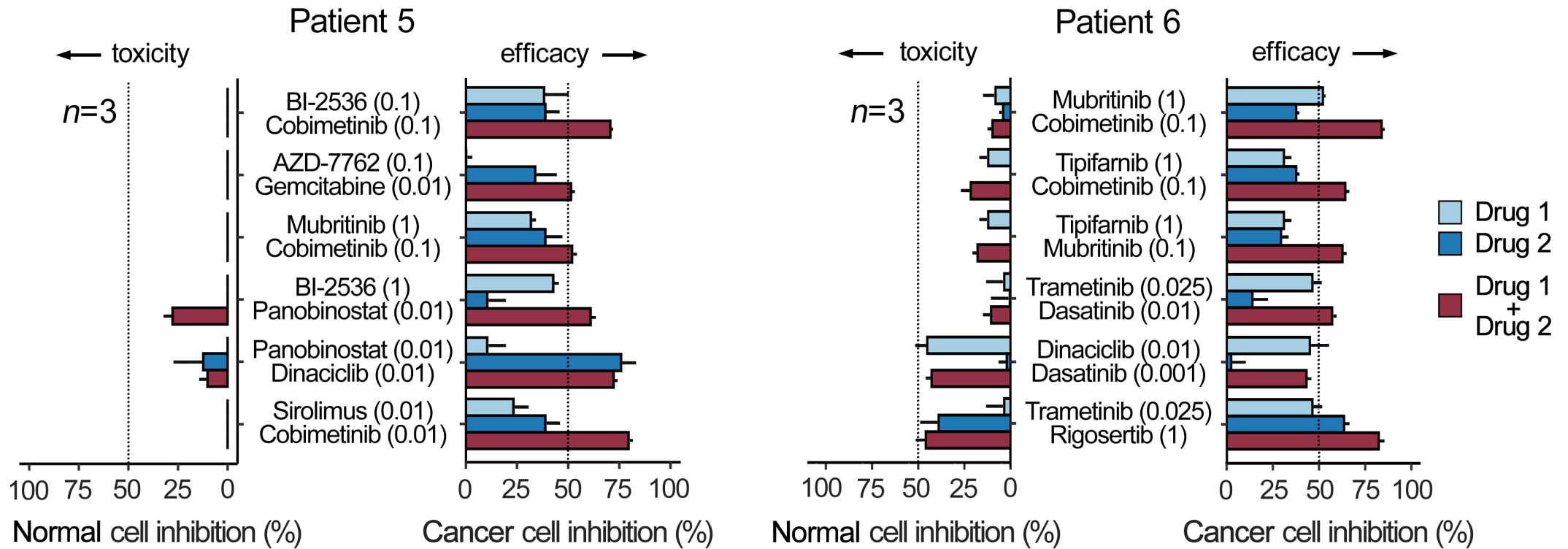


Analysis



**blast cells vs.
lymphoid cells**

Trade-off between co-inhibition of healthy and cancer cells

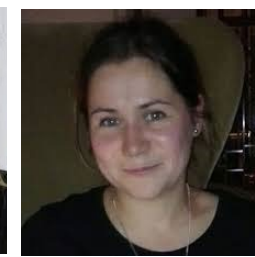


Low toxicity: <50% relative co-inhibition of lymphoid cells (healthy cells)

Application to ovarian cancer patients Validation in patient-derived organoids



Kyriaki Driva



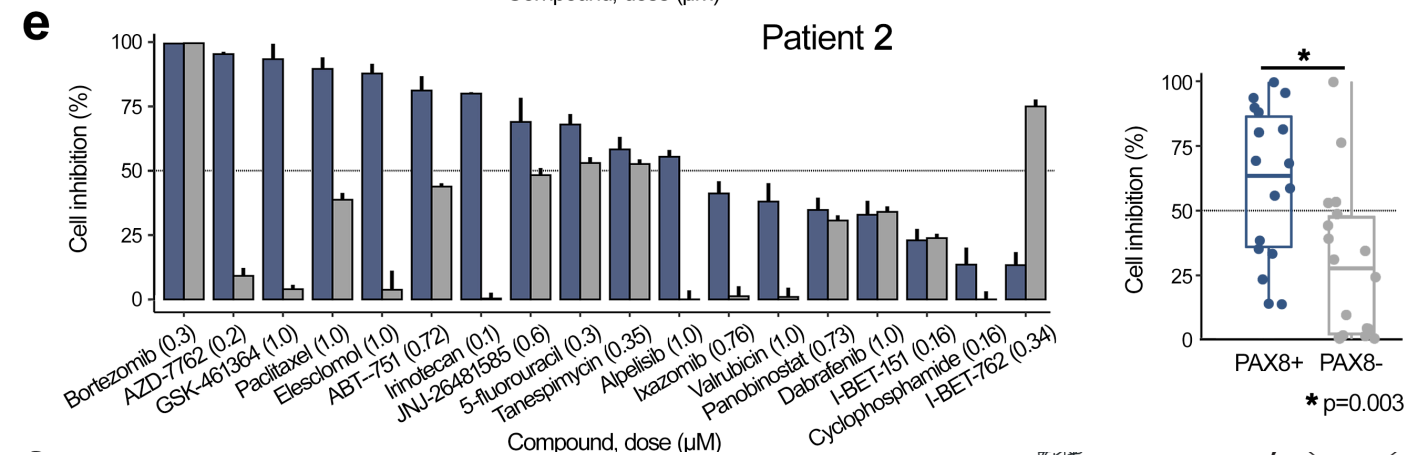
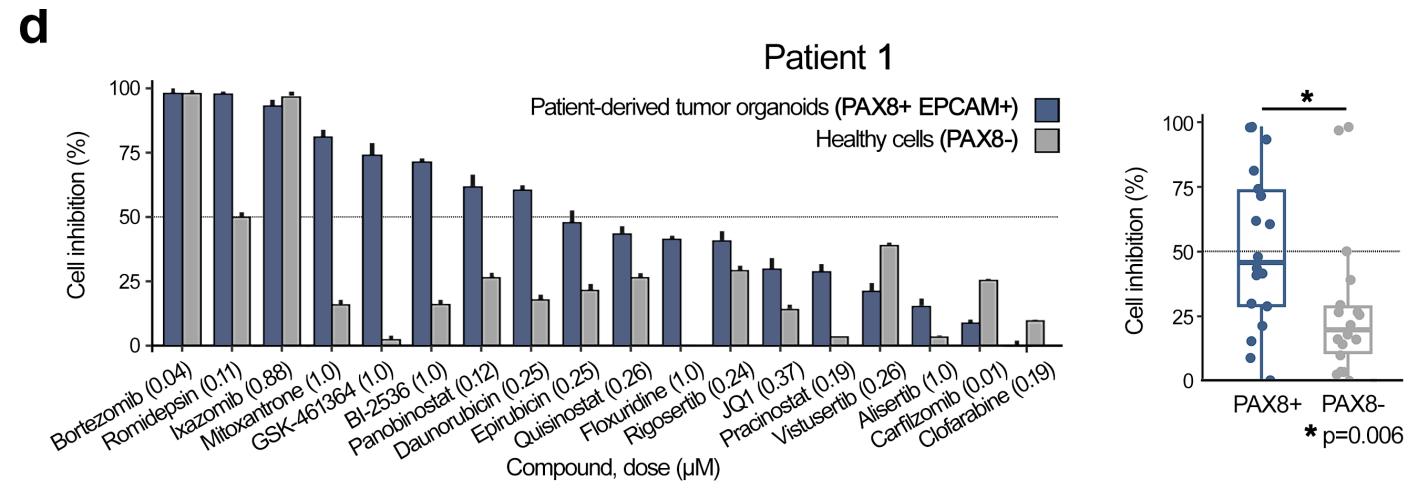
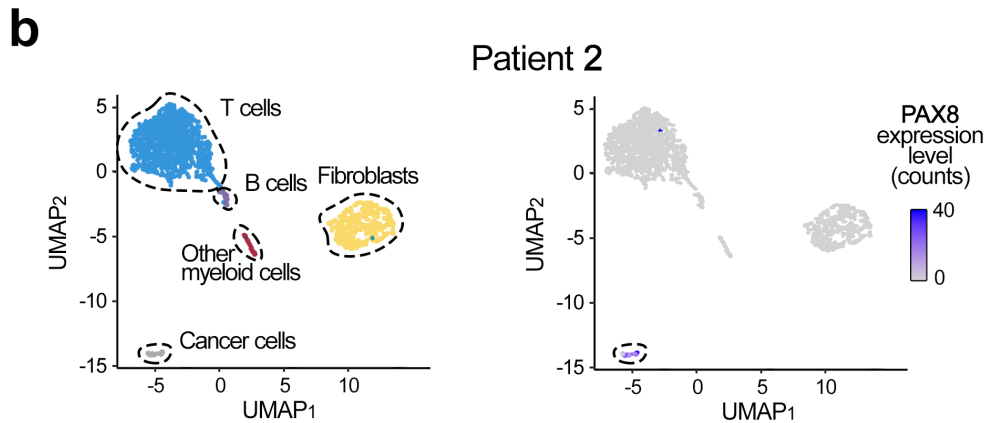
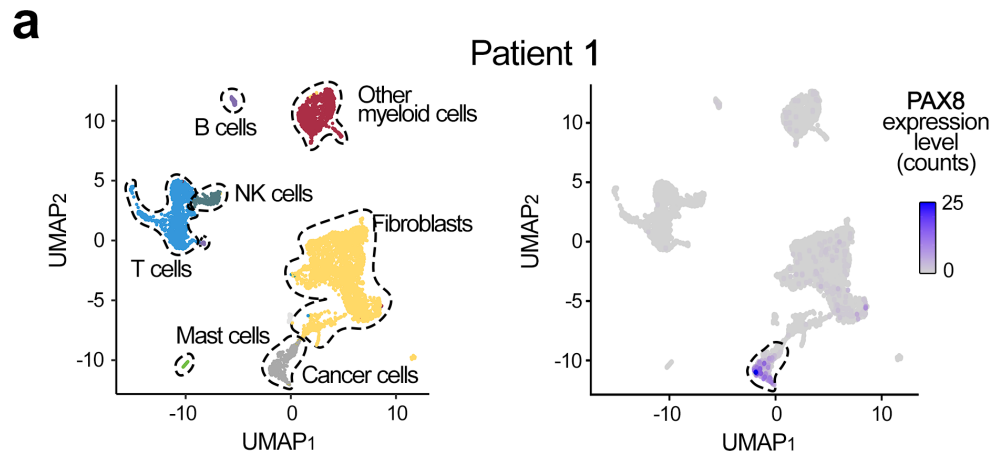
Daria Bulanova



Wojciech Senkowski



Lidia M. Galceran



Conclusions

- Drug combination predictions using **only scRNA-seq** from patient samples
- Fast means for identifying **targeted treatments and doses** at subclone and patient level (diagnosis vs. relapse/refractory)
- Can be applied for tumours that are not easily amenable to *ex vivo* drug testing (most solid tumours)
- **96%** of the multi-targeting treatments were confirmed to exhibit selective synergy and **86%** low toxicity to normal cells
- Potential for improved therapeutic efficacy and safety
- Challenging to identify biomarkers from scRNA-seq data

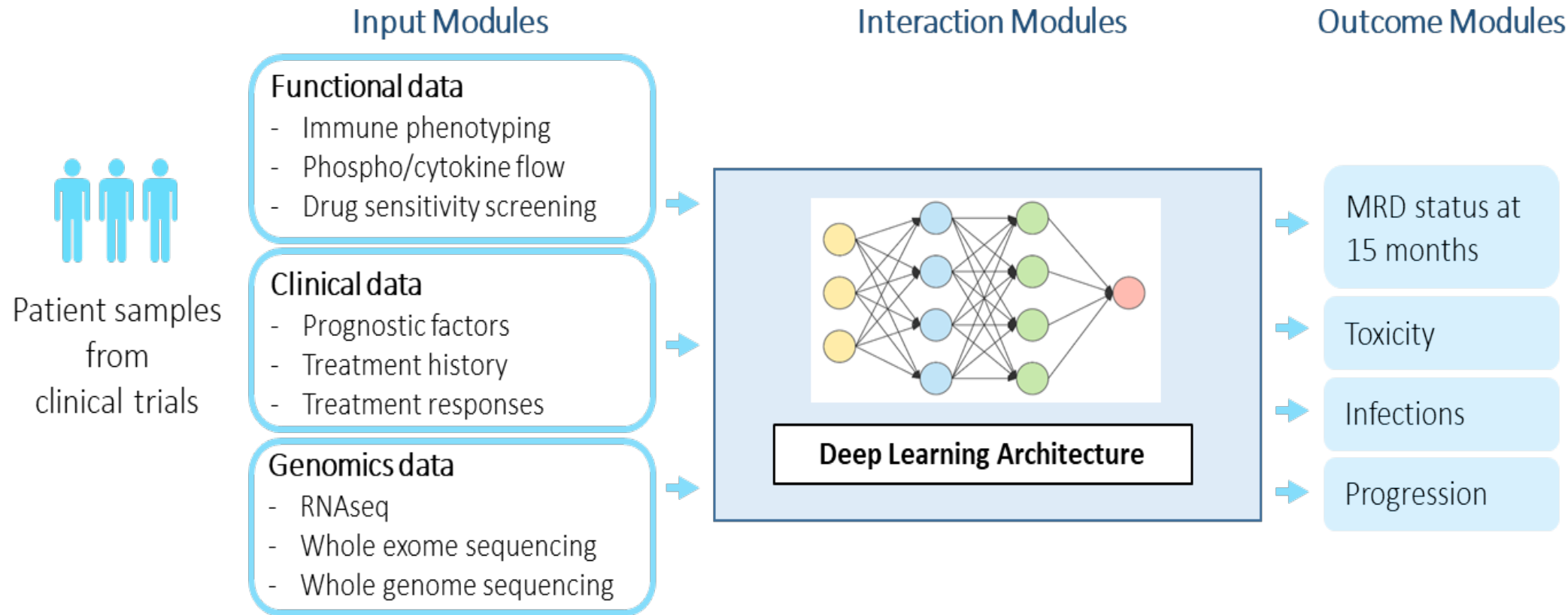
Systematic approach to predict effective drug dose-combinations that minimize toxicity to healthy cells while selectively co-inhibit cancer cells
➔ **clinical response prediction as the next step (VenEx trial)**





ERA PerMed CLL-CLUE project

AI-based treatment decision support system



Rigshospitalet



UNIKLINIK
KÖLN



Oslo
University Hospital

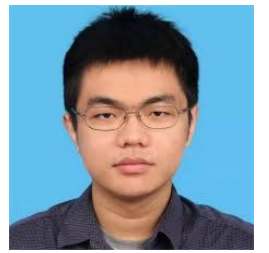


University of
Zurich^{UZH}

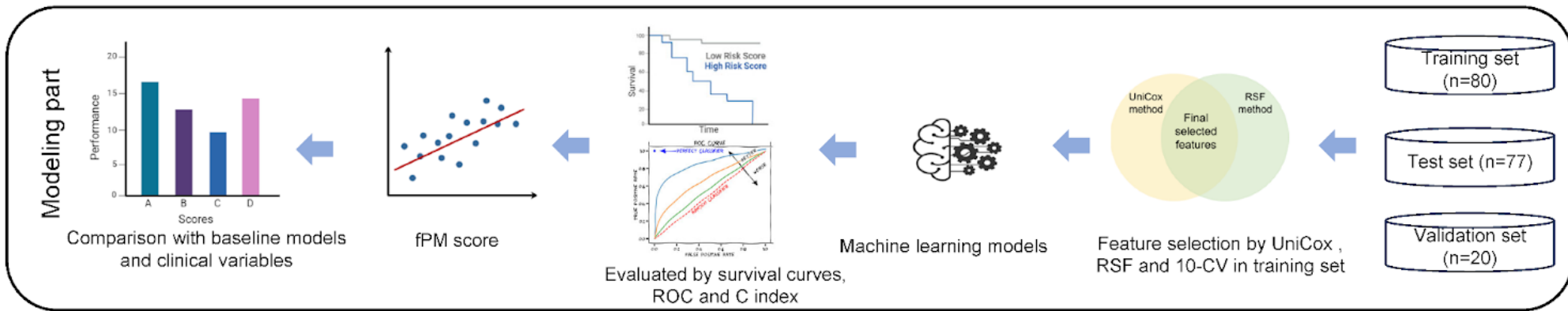
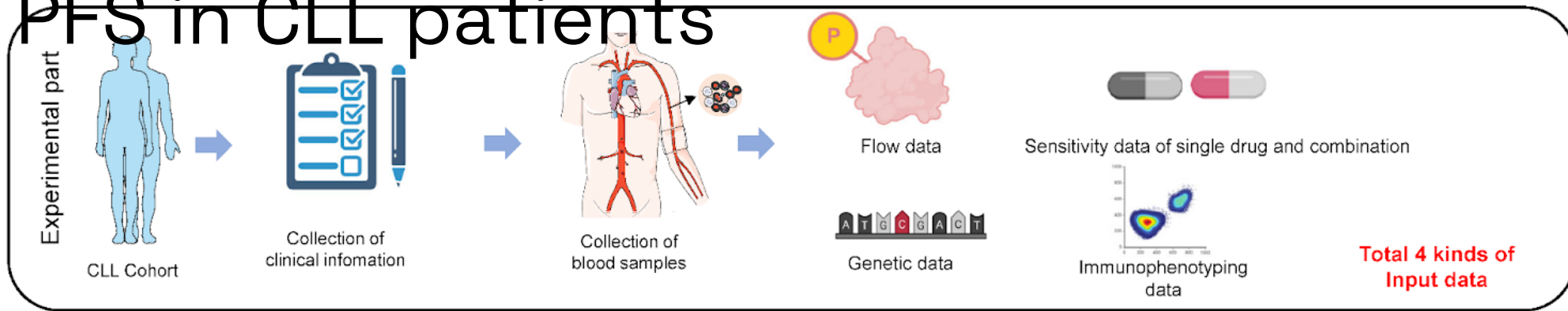
syreon
Research Romania

FIMM
Institute for Molecular Medicine Finland
Nordic EMBL Partnership for Molecular Medicine

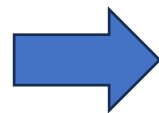
Functional precision medicine (fPM) score for prognostic predictions of PFS in CLL patients



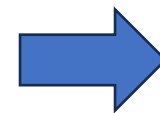
Weikaixin Kong (FIMM)



Clinical expertise



Profiling technology



Machine learning

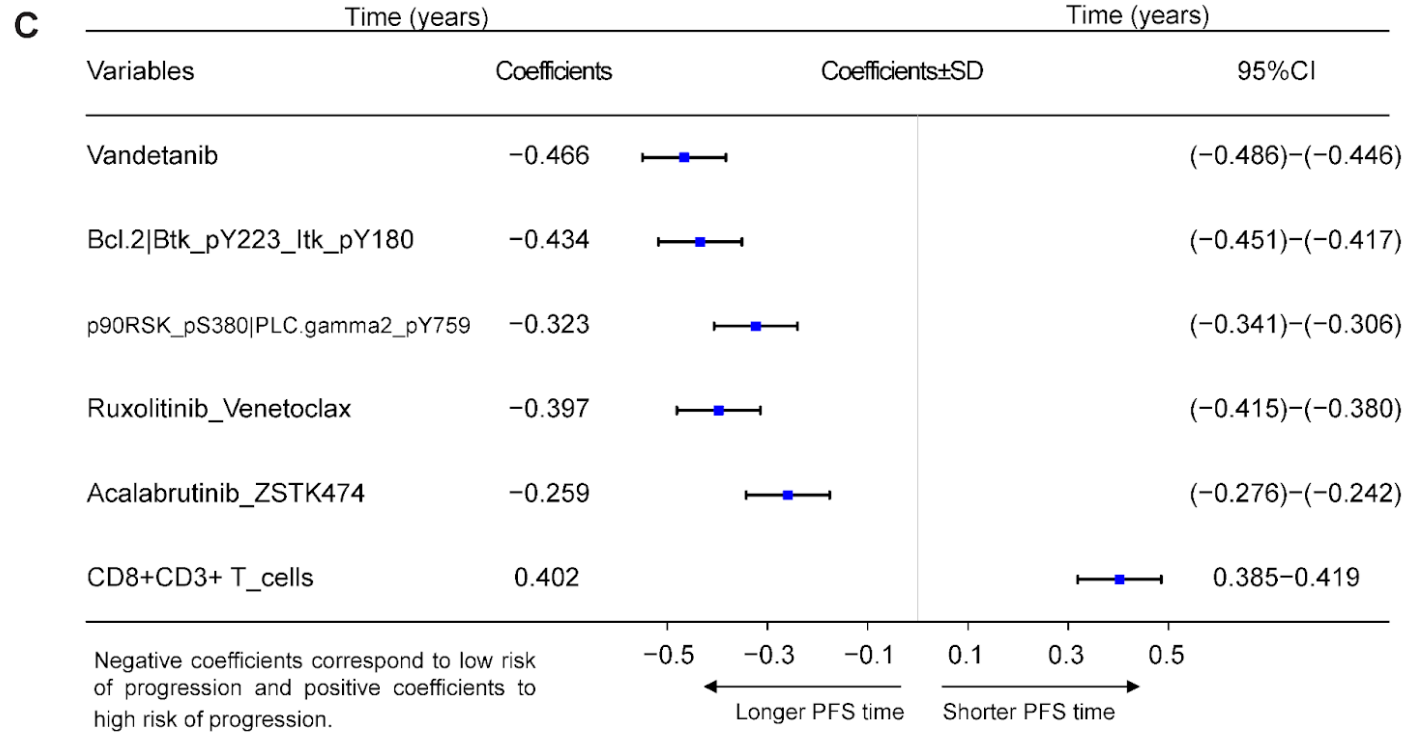
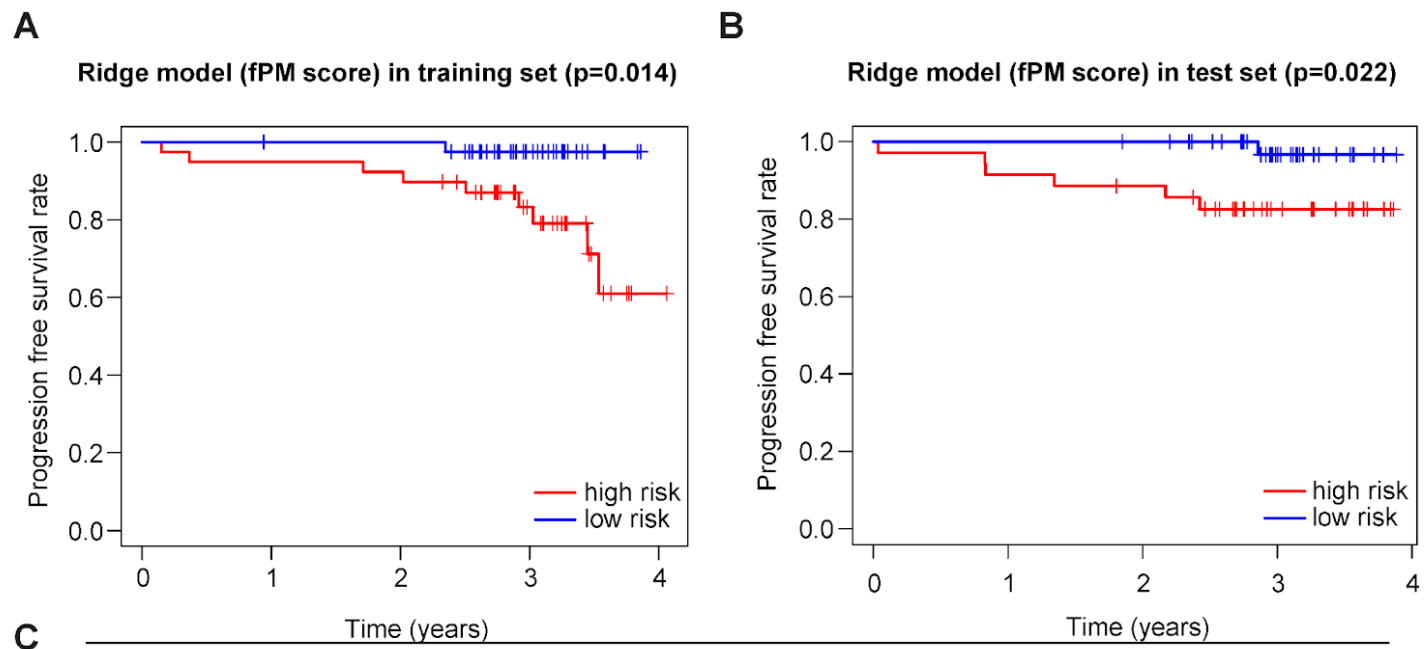


Figure 1. Survival curves of the fPM high and low-risk groups in **(A)** training cohort (n=80) and **(B)** test cohort (n=77). **(C)** The coefficients of the variables of the fPM score estimated in the training cohort. The median value of the fPM score in the training set (-0.01687) was used as the cutoff to separate between the high-risk and the low-risk patient groups. Statistical significance of the PFS differences between the two groups was assessed with the log rank test.

Personalized Cancer Medicine

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Satu Mustjoki

Kimmo Porkka

Mika Kontro

Erkki Elonen

Hanna Koskela

Mette Ilander

Emma Anderson

Paavo Pietarinen

Jaakko Vartia

Minna Lehto

Mervi Saari

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AML case study

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Mika Kontro
Kimmo Porkka
Caroline A. Heckman
Nemo Ikonen
Philipp Sergeev
Juho Miettinen
Imre Västrik
Esa Pitkänen

HGSC case study

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Wojciech Senkowski
Laura Gall-Mas
Lidia Moyano-Galceran
Anna Vähärautio
Krister Wennerberg

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the patients and their
families for participating
in the two studies